

On the Existence of Structurally Different Isobaric B_n Fragment Ions

Touradj Solouki, Behrooz Zekavat, and Mahsan Miladi

Department of Chemistry, 5706 Aubert Hall, University of Maine, Orono, ME 04469-5706

INTRODUCTION

In 2001, we reported the existence of structurally different isobaric b fragments for WHYQL [1]. Recently, we reported on the details of the hydrogen/deuterium (H/D) exchange reactions for structurally different isobaric singly [2] and doubly-charged b_n fragment ions [3] of various peptides during gas-phase fragmentation. HD exchange ion-molecule reactions were used to demonstrate the presence of at least two different isomer-conformers reactions [2]. In other words, HD exchange reactions of isolated single isomers (¹²C₁₀₀) of b_n fragment ions yielded bimodal distribution patterns and allowed for observation of the two resolved ion populations. Production of structurally different b_n fragment ions can play a major role in peptide/protein fragmentation and sequencing (i.e., sequence scrambling [4]). Presently several research groups around the world, recognizing the importance of our initial observations and the significance of different b-type fragment ions in peptide/protein sequencing, are performing amino acid residue dependent and other systematic studies to explore the details of such fragmentation mechanisms. In this study, for the first time, we report the influence of other structural (e.g., N-terminus acetylation) and experimental parameters (e.g., ion fragmentation energetic) on the production of structurally different b-type fragment ions; we also report on the use of peptide ion gas phase basicity (GB) measurements in this context. Based on our findings, we propose the use of standard peptides and b fragment ions (as molecular-ion thermometers) to reduce potential non-stoichiometric experimental variations and enhance meaningful data comparison.

GOALS

Our goal is to utilize (1) gas-phase proton transfer reactions, (1) collision induced dissociation (CID), and electron capture dissociation (ECD) to examine the existence of isobaric singly- and doubly-charged b_n fragment ions. We are interested in the mechanisms and pathways involved in the formation of different isobaric b fragment ions. Here, we explore the influence of structural variations (e.g., effect of N-terminus acetylation of model peptides) and experimental conditions on the appearance or disappearance of structurally different b fragment ions.

EXPERIMENTAL

Instrumentation: ESI/FT-ICR mass spectra were acquired with an FT-ICR mass spectrometer equipped with an open-ended cylindrical Penning trap (Thermo IonPep Corp., now a division of Varian, Inc., Lake Forest, CA) and a 9.4 Tesla superconducting magnet (Cryoconcepts Inc., Oak Ridge, TN). Ions were generated externally using an Analytica source (Analytica of Branford Inc., Branford, CT) equipped with in-house built spraying setup. Peptide fragment ions were generated using nozzle-skimmer fragmentation technique. A pulsed-deck valve was used for controlled introduction of the reagent gas. The GB measurements and/or HD exchange reactions into the ICR cell. Stored waveform inverse FT (SWIFT) techniques were utilized for ion isolation [5].

Sample Preparation: All of the chemical solvents and reagents, and peptides were purchased from commercial sources and used without any further purification except for applying conventional freeze-dry cycles as deemed necessary (e.g., for reagents used in proton transfer reactions). Micromolar concentration of individual or mixture of peptide solutions in methanol/water: acetic acid (48.5:49.5:1) were used for the ESI experiments.

Data Analysis: Normalized ion intensities were calculated from I_i/I_j where I_i is the ion intensity (i.e., ¹²C₁₀₀ peak intensity) and I_j is the total ion intensity (i.e., ¹²C₁₀₀ + ¹³C₁₀₀ + ... in HDX and ¹²C₁₀₀ + ¹³C₁₀₀ + ... in proton transfer reactions). To calculate the rate constants, an exponential curve fitting (using Origin 7.0 (OriginLab Corporation, MA)) was performed to extract the time constants (τ) for "fast" and "slow" populations. Thus the rate constants were estimated using the following equation:

$$k = 1/(\tau \times \ln 2)$$

k: rate constant (cm³ Mol⁻¹ s⁻¹)
 τ: time constant (s)
 [N]: number density of the neutral reagent (Mol cm⁻³)

RESULTS AND DISCUSSION

(a) Proton Affinities of Different Structures of b₁₀²⁺ Fragment Ion

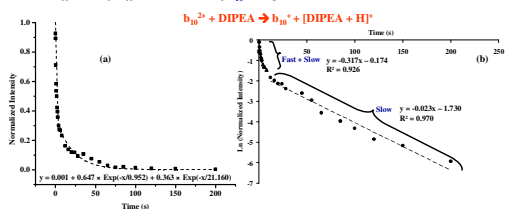


Table 1. CID products of b₁₀²⁺ fragment ion.

Fragment type	Rate Constant (cm ³ Mol ⁻¹ s ⁻¹)	GB (kcal mol ⁻¹)
b ₁₀ ²⁺ (fast)	4.1 (± 0.3) × 10 ⁻¹⁰	229.29 (± 0.06)
b ₁₀ ²⁺ (slow)	1.8 (± 0.4) × 10 ⁻¹¹	227.30 (± 0.13)

Figure 2. (a) Temporal and (b) semi-log plots of normalized ion intensity vs. proton transfer reaction time for the disappearance of [b₁₀(¹²C₁₀₀)]²⁺. Neutral reagent: diisopropylethylamine (DIPEA) (PA = 237.6 kcal/mol), P(DIPEA) = 7.5 × 10⁻⁹ torr. The calculated rate constants and gas phase basicity of fast and slow proton transferring populations are shown in Table 1.

(b) Collision Induced Dissociation (CID) Mass Spectrum of b₁₀²⁺ Fragment Ion

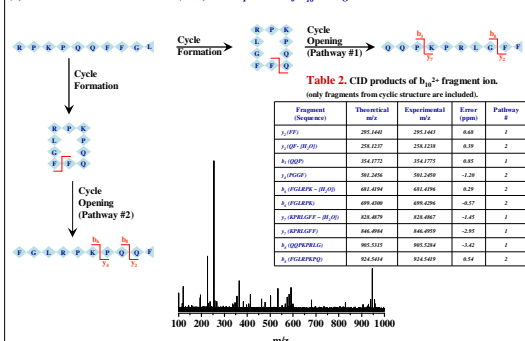


Table 2. CID products of b₁₀²⁺ fragment ion. (only fragments from cyclic structure are included).

Fragment (Sequence)	Theoretical m/z	Experimental m/z	Error (ppm)	Relative Intensity
b ₁₀ (PP) ²⁺	295.1444	295.1443	0.49	1
b ₁₀ (PP) ²⁺	298.1227	298.1228	0.39	2
b ₁₀ (PP) ²⁺	301.1010	301.1011	0.49	1
b ₁₀ (PP) ²⁺	304.0793	304.0794	0.28	2
b ₁₀ (PP) ²⁺	307.0576	307.0576	0.29	2
b ₁₀ (PP) ²⁺	310.0359	310.0359	0.27	2
b ₁₀ (PP) ²⁺	313.0142	313.0142	0.45	1
b ₁₀ (PP) ²⁺	315.9925	315.9925	0.31	1
b ₁₀ (PP) ²⁺	318.9708	318.9708	0.42	1
b ₁₀ (PP) ²⁺	321.9491	321.9491	0.54	2

Figure 2. CID/FT-ICR mass spectrum of b₁₀²⁺ fragment ion illustrating the indirect fragment ion formation from cyclic structure (assignments of the fragments with theoretical and experimental m/z) are shown in Table 2).

Note: The CID spectrum in Figure 2 shows the potential for cyclic structure formation for b₁₀²⁺ fragment ions and "sequence scrambling" in subsequent fragmentation (e.g., MS²).

(c) Electron Capture Dissociation (ECD) Mass Spectrum of b₁₀²⁺ Fragment Ion

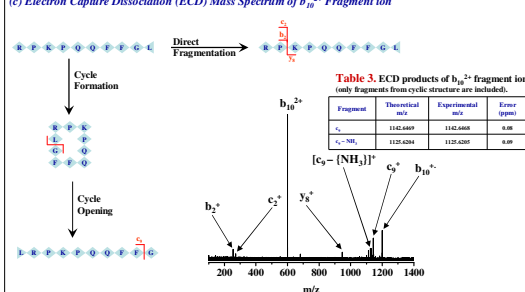


Table 3. ECD products of b₁₀²⁺ fragment ion (only fragments from cyclic structure are included).

Fragment	Theoretical m/z	Experimental m/z	Error (ppm)
b ₁₀ ²⁺	1102.4469	1102.4468	0.38
c ₁₀ -NH ₂	1125.4254	1125.4255	0.49

Figure 3. ECD/FT-ICR mass spectrum of b₁₀²⁺ fragment ion showing the direct and indirect fragment ion formation from cyclic and linear structures of b₁₀²⁺, respectively (assignments of the fragments (with theoretical and experimental m/z) are shown in Table 3).

Note: The ECD spectrum in Figure 3 shows the evidence for potential cyclic b₁₀²⁺ structure formation.

(d) Possibility for interconversion between "Fast" and "Slow" b_n Fragment Ions

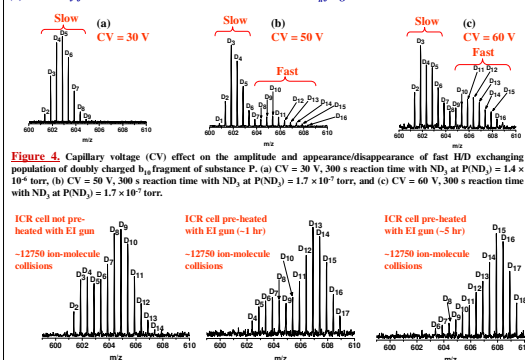


Figure 4. Capillary voltage (CV) effect on the amplitude and appearance/disappearance of fast HD exchanging population of doubly charged b₁₀ fragment of substance P. (a) CV = 30 V, 300 s reaction time with ND₃ at P(ND₃) = 1.4 × 10⁻⁸ torr, (b) CV = 50 V, 300 s reaction time with ND₃ at P(ND₃) = 1.7 × 10⁻⁷ torr, and (c) CV = 60 V, 300 s reaction time with ND₃ at P(ND₃) = 1.7 × 10⁻⁷ torr.

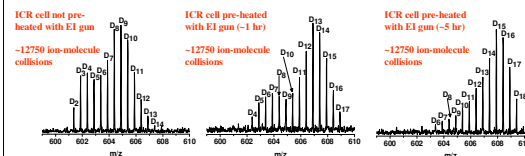


Figure 5. Temperature effect (using ICR cell electron emission (EI) gun) on the amplitude and disappearance of fast HD exchanging population of doubly charged b₁₀ fragment of substance P. (a) EI gun "off", (b) EI gun "on" -1 hr, and (c) EI gun "on" -5 hr. ¹³C₁₀₀ isotopic peak of b₁₀²⁺ was SWIFT isolated and then reacted with ND₃ for 500 s at P(ND₃) = 5.0 × 10⁻⁹ torr.

(e) Effect of Peptide N-terminus Acetylation on the b-type Fragment Structure

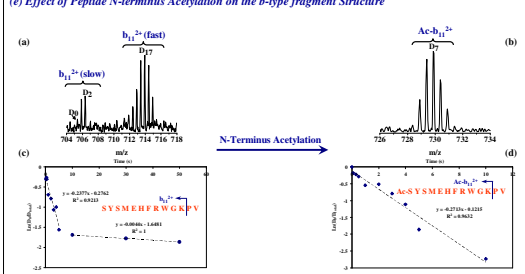


Figure 6. ESI/FT-ICR mass spectrum of SWIFT-isolated (a) b₁₁²⁺ of des-Ac-a-Melanocyte and (b) acetylated b₁₁²⁺ of a-Melanocyte (i.e., Ac-b₁₁²⁺) after 300 s H/D exchange with ND₃ at P(ND₃) = 1.1 × 10⁻⁷ torr. The semi-log plots for disappearance of D₀ ion population of b₁₁²⁺ and Ac-b₁₁²⁺ are shown in Figures 1c and 1d, respectively.

Note: While b₁₁²⁺ fragment ion shows at least two structures with different reactivities in HD exchange reaction, Ac-b₁₁²⁺ fragment ion shows a single structure. The results in Figure 6 suggest that N-terminus acetylation potentially prevents the cyclization of b₁₁²⁺ fragment ion.

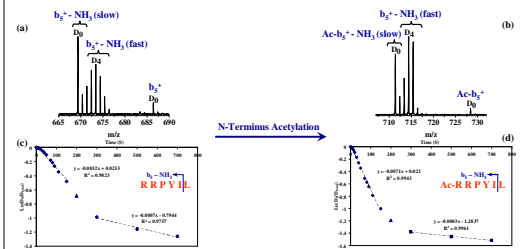


Figure 7. ESI/FT-ICR mass spectrum of SWIFT-isolated (a) b₁^{*} and b₁^{*}-NH₂ of Neurotensin (b) N-terminus acetylated b₁^{*} (i.e., Ac-b₁^{*}) and b₁^{*}-NH₂ of Ac-Neurotensin after 700 s H/D exchange with ND₃ at P(ND₃) = 1.5 × 10⁻⁹ torr. The semi-log plots for disappearance of D₀ ion population of b₁^{*}-NH₂ and Ac-b₁^{*}-NH₂ are shown in Figures 2c and 2d, respectively.

Note: While neither b₁^{*} nor Ac-b₁^{*} fragment ions show HD exchange reactivity, loss of NH₂ from both fragments (i.e., b₁^{*}-NH₂ and Ac-b₁^{*}-NH₂) activates HD exchange reaction channels.

CONCLUSIONS

- Experimental gas phase basicity (GB) values for structurally different isobaric b_n²⁺ fragment ions of substance P showed ~2 kcal/mol GB difference.
- CID and ECD results suggest the presence of a cyclic structure for b₁₀²⁺ fragment ion of substance P.
- The preliminary results suggest that the relative abundances of "fast" and "slow" HD exchanging ion populations can be controlled in the ion source (i.e., metal capillary voltage effect) or in the ICR cell (i.e., temperature effects). Although the influence of temperature variations on HD exchange rate must be studied in detail to account for its contribution.
- N-terminus acetylation of des-Ac-a-melanocyte prevented the formation of "fast" HD exchanging b₁₁²⁺ ion population, contrary to the presence of two structures for the non-acetylated b₁₁²⁺ fragment.
- Neither b₁^{*} nor Ac-b₁^{*} fragment ions of neurotensin show HD exchange reactivities; however, b₁^{*}-NH₂ and Ac-b₁^{*}-NH₂ yield at least two structures with different HD exchange reactivities/patterns.
- The presence of different isomer/conformer b fragment ions is not limited to small peptides. This is a significant finding and has very important consequences in all proteomics studies and gas-phase protein sequencing. To reduce experimental variabilities, we propose the use of standard peptides and b fragment ions (as molecular-ion thermometers) in all systematic studies involving the formation of different b fragment ions.
- Future experimental and theoretical calculations should provide additional details on the influence of various thermochemical parameters on fragmentation mechanisms.

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