

Memory and Epigenetics: Role of Estrogen

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Introduction

Evidence that hormones could regulate behavior was first demonstrated in 1849 when Arnold Berthold showed that castration and reimplantation of the testes in roosters affected normal male development, sexual desire, and aggression. However, only in the last 30 years has it been demonstrated that sex steroid hormones are poised to mediate memory formation. Some of the first evidence supporting this notion came from the discovery of estrogen receptors in the dorsal hippocampus and the entorhinal cortex (Loy et al., 1988; Maggi et al., 1989) and from seminal work demonstrating that spine density in the CA1 region of the hippocampus could be regulated by estrogens, including the potent 17 β -estradiol (E₂), and progesterone in female rats (Gould et al., 1990; Woolley et al., 1990; Woolley and McEwen, 1992, 1993). In the nearly three decades since the initial demonstration that ovarian hormones regulate CA1 dendritic spine density, preclinical research has provided evidence that E₂ can enhance learning and memory in adults of a variety of species, including songbirds, rodents, nonhuman primates, and humans (for reviews see Maki, 2012; Hammond and Gibbs, 2011; Frick, 2009, 2012; Schlinger and Remage-Healey, 2012; Bimonte-Nelson et al., 2010). Recent work has demonstrated that E₂ employs numerous cellular and molecular mechanisms, including epigenetic processes, to influence memory formation, and these epigenetic mechanisms will constitute the focus of this review. A general overview of the beneficial effects of E₂ on memory will first be described below, followed by a discussion of the epigenetic mechanisms through which E₂ exerts these effects.

The Role of E₂ in Cognitive Function

Estradiol and Cognitive Function in Humans

Clinical studies investigating the estrogenic regulation of memory have focused largely on the effects of estrogens in the context of aging (i.e., as it relates to menopause), or changes in performance in cognitive tasks across the menstrual cycle. With respect to aging, numerous studies have shown that ovarian hormone loss due to natural or surgical menopause impairs various aspects of cognitive function, including verbal and spatial memory (Sherwin and Henry, 2008) (Fig. 1). Menopausal women are also at increased risk of Alzheimer's disease relative to men, even when accounting for women's longer lifespans (Dye et al., 2012; Launer et al., 1999). Indeed, longer periods of lifetime estrogen exposure are significantly and negatively correlated with the likelihood of developing Alzheimer's disease in women (Fox et al., 2013). The deleterious effects of menopause on cognition and risk of Alzheimer's disease can be reduced by E₂ therapy (Rocca et al., 2007, 2011), suggesting not only that E₂ loss at menopause contributes to memory dysfunction, but also that maintaining E₂ levels at menopause may help prevent Alzheimer's disease and reduce age-related cognitive decline.

Evidence also indicates that E₂ fluctuations across the month-long menstrual cycle in premenopausal women impacts certain aspects of cognitive function. For example, women experiencing high E₂ levels during the midluteal phase of the cycle perform better in tests of verbal fluency, fine motor, and perceptual speed than women experiencing low E₂ levels during the menstrual

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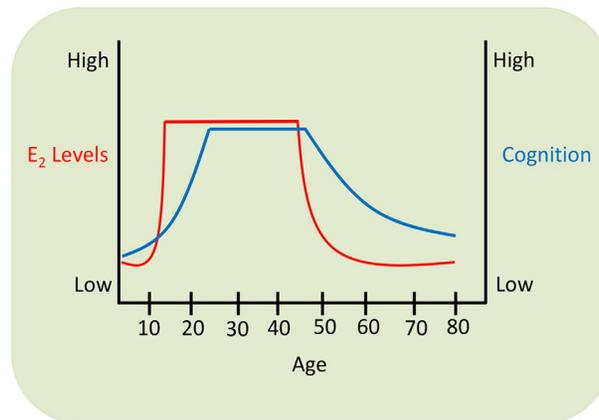


Fig. 1 Schematic illustration of changes in estradiol (E_2) levels and cognitive abilities across the life span in women. E_2 levels surge upwards at puberty and remain elevated (although fluctuating across the menstrual cycle) during the reproductive years. In the early 50's, E_2 levels drop substantially at menopause. Cognitive function reaches adult levels in the early-mid 20s and then remains relatively stable until middle-age, at which point certain aspects of cognitive function (e.g., episodic memory, divided attention, spatial navigation) decline gradually with age.

phase (Hampson, 1990; Maki et al., 2002). Some evidence suggests that visual memory is also poorer during the menstrual phase (Phillips and Sherwin, 1992). High levels of E_2 do not always correlate with better performance on cognitive tasks, however, as women's performance appears to be best in tasks in which men traditionally outperform females (e.g., those related to spatial ability) when E_2 levels are low (Hampson, 1990; Maki et al., 2002).

In addition to its role in regulating normal cognitive function, E_2 has also been implicated in several disorders that disproportionately affect women relative to men, including Alzheimer's disease, posttraumatic stress disorder (PTSD), and certain aspects of addiction. Although beyond the scope of this article, reviews on the role of E_2 in Alzheimer's disease (Dye et al., 2012), PTSD (Albert et al., 2015; Milad et al., 2010), traumatic brain injury (Asl et al., 2013; Day et al., 2013b), and substance abuse (Becker, 2016) provide important insights into the ways in which sex-steroid hormones may impact disorders characterized by memory dysregulation, and suggest directions for future research to better understand underlying neural mechanisms.

Animal Models Demonstrate the Importance of E_2 in Cognitive Function

Basic scientific research conducted in animal models is critical for mechanistic investigations into the role of E_2 in cognitive function. E_2 -mediated actions in brain regions that support cognition, such as the hippocampus, are complex and impacted by numerous factors, including dose, duration of treatment, age, length of ovarian hormone deprivation prior to treatment, type of cognitive task, timing of administration relative to testing, task difficulty, and reproductive history (Acosta et al., 2009, 2010; Luine, 2014; Daniel, 2006; Frick, 2009). However, some generalizations can be drawn, particularly among studies utilizing rodent models. The majority of rodent studies support the conclusion that E_2 facilitates learning and memory in behavioral tasks that require the hippocampus (Packard and Teather, 1997; Fader et al., 1998; Daniel et al., 1999; Luine et al., 2003; Walf et al., 2008; Fernandez et al., 2008; Lewis et al., 2008; Zhao et al., 2010; Daniel, 2006; Fan et al., 2010); see (Daniel, 2006; Tuscher et al., 2015) for review. For example, young adult female rodents treated with exogenous E_2 exhibit enhanced spatial memory in the object placement, Morris water maze, radial arm maze, and T-maze tasks (Sandstrom and Williams, 2004; Luine et al., 1998; Fader et al., 1998, 1999; Daniel et al., 1997; Bowman et al., 2002; Bimonte et al., 2002). E_2 can also facilitate memory in a number of nonspatial tasks, as demonstrated in object recognition (Fernandez et al., 2008; Fortress et al., 2013; Boulware et al., 2013), social recognition (Phan et al., 2012), inhibitory avoidance (Singh et al., 1994; Rhodes and Frye, 2004), fear conditioning (Lebron-Milad and Milad, 2012; Chang et al., 2009; Barha et al., 2010; Zeidan et al., 2011; Milad et al., 2010), and trace eyeblink conditioning (Leuner et al., 2004). Collectively, these studies provide evidence that E_2 treatment can benefit hippocampal memory in numerous behavioral tasks. The molecular mechanisms through which E_2 exerts these beneficial effects will be discussed in greater detail below.

Estrogen Receptor Expression and Cell-Signaling Mechanisms

Protein synthesis is an essential component of memory formation, and E_2 regulates the synthesis of new proteins through at least two different estrogen receptor (ER)-mediated mechanisms: the classical genomic pathway and the rapid nonclassical activation of cell-signaling pathways. Many of the brain regions that support memory formation express classical intracellular ERs ($ER\alpha$ and $ER\beta$), which are found within the cytoplasm and nucleus of the cell. Both $ER\alpha$ and $ER\beta$ have their own distinct patterns of expression in the cerebral cortex, basal forebrain, amygdala, prefrontal cortex, and hippocampus in a variety of species, including mouse, rat, nonhuman primates, and humans (Gillies and McArthur, 2010). The classical "genomic" action of E_2 is initiated once the hormone diffuses through the target cell's outer membrane and binds $ER\alpha$ or $ER\beta$ within the cytoplasm (Nelson, 2000). Once the E_2 -ER complex is formed, it translocates to the nucleus, where it binds to estrogen response elements on the DNA. Here, the complex acts

as a transcription factor, and can initiate the transcription of E_2 -sensitive genes that help maintain the neural circuitry that ultimately influences behavior (Heldring et al., 2007; Jensen, 1962). Changes in gene expression elicited by such nuclear ER-hormone interactions via the genomic mode of action occur slowly (on the scale of hours—days) and are thought to yield long lasting changes.

E_2 can also influence cell function in a nonclassical manner by binding to membrane-associated ERs (mERs; e.g., GPER, Gq-mER; Srivastava and Evans, 2013), classical ERs located near the membrane (Boulware et al., 2005, 2013), or by interacting with neurotransmitter receptors (e.g., mGluRs, NMDARs; Lewis et al., 2008; Boulware et al., 2005, 2013) to rapidly activate intracellular signaling pathways on the order of seconds to minutes (Gillies and McArthur, 2010). Although these mechanisms are often referred to as “nongenomic,” this designation should not be taken literally, as activation of mERs can ultimately influence gene transcription. Rather, it should be thought of as way to distinguish between the effects of mERs and classical nuclear ER activation.

Work in our own lab has focused on the cell-signaling pathways through which E_2 regulates function in the dorsal hippocampus (DH) and medial prefrontal cortex, brain regions critical for learning and memory and whose function is compromised in various neuropsychiatric disorders and during aging (Small et al., 2011; Maillet and Rajah, 2013; Sampath et al., 2017). Rapid activation of cell-signaling cascades in these regions allows for acute modulation of cellular function in response to an experience (i.e., learning). Several pathways previously identified in studies focused on memory modulation in male rodents, such as extracellular signal-regulated kinase/mitogen activated protein kinase (ERK/MAPK), phosphatidylinositol 3-kinase (PI3K), protein kinase A (PKA), calcium calmodulin kinase II (CaMKII), and mammalian target of rapamycin (mTOR) (Adams and Sweatt, 2002; Horwood et al., 2006; Impey et al., 1998; Silva et al., 1992; Hoeffler and Klann, 2010) are also regulated by E_2 in ovariectomized females. E_2 -mediated activation of these pathways has been linked to changes in neuronal excitability (Woolley, 2007), long-term potentiation (LTP) (Smith and McMahon, 2005), spinogenesis (Tuscher et al., 2016a), and memory enhancement (Fernandez et al., 2008; Fan et al., 2010; Fortress et al., 2013) in ovariectomized females. Specifically, E_2 infusion directly into the DH rapidly activates cell-signaling cascades like ERK, PI3K, PKA and mTOR within 5 min of DH infusion (Fernandez et al., 2008; Fan et al., 2010; Fortress et al., 2013). Further, activation of ERK, PI3K, PKA, and mTOR are all required for the beneficial mnemonic effects of E_2 in hippocampus-dependent tasks, as pharmacological inhibition of any of these pathways prevents the memory-enhancing effects of E_2 in ovariectomized female mice (Fernandez et al., 2008; Fan et al., 2010; Fortress et al., 2013) (Fig. 2). As will be discussed below, E_2 -mediated activation of certain pathways, such as ERK, can also alter gene transcription by regulating downstream epigenetic modifications such as DNA methylation and histone acetylation. Before addressing these data, the next section will first review the major epigenetic mechanisms that regulate hippocampal memory.

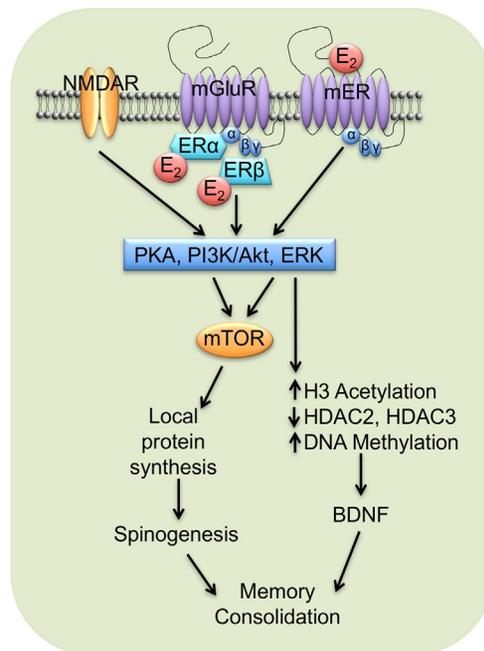


Fig. 2 Schematic representation of the cellular mechanisms through which E_2 regulates memory in ovariectomized mice. In the dorsal hippocampus, E_2 interacts with intracellular ERs (ER α and ER β), mERs, and neurotransmitter receptors to rapidly activate PKA, PI3K/Akt, ERK, and mTOR cell signaling, which then triggers spinogenesis via local protein synthesis. Similarly, phosphorylation of ERK can enhance gene transcription via alterations in H3 acetylation, HDAC protein levels, and DNA methylation. These changes all contribute to the memory-enhancing effects of acute dorsal hippocampal E_2 infusion on memory consolidation. Abbreviations: NMDAR, N-methyl-D-aspartate receptor; mGluR, metabotropic glutamate receptor; mER, membrane estrogen receptor; PKA, protein kinase A; PI3K, phosphatidylinositol 3-kinase; mTOR, mammalian target of rapamycin; H3, histone 3; HDAC, histone deacetylase; BDNF, brain derived neurotrophic factor.

Epigenetic Modifications Related to Synaptic Plasticity and Memory

The term “epigenetics” refers to the idea that the expression of genes can be regulated “above the gene” by altering accessibility to genes rather than the genetic code itself (Crick, 1984; Holliday, 1999). Chromosomes are comprised of DNA tightly coiled around nucleosome subunits. Each nucleosome consists of an octamer containing eight histone (H) proteins, two each of H2A, H2B, H3, and H4 (Fig. 3). Each histone protein has a N-terminal tail that can be altered through posttranslational modifications such as acetylation, methylation, phosphorylation, ubiquitination, and SUMOylation (Rothbart and Strahl, 2014) (Fig. 3). Chromatin is formed from the supercoiling of adjacent nucleosomes which are connected by the linker protein H1 (Mazzio and Soliman, 2012). The ability of a certain segment of chromatin to be modified depends on the posttranslational modification state of its histone tail, which controls how tightly wound the chromatin structure is and, therefore, whether the 3D structure is permissive to binding factors that regulate transcription. Each major modification and its individual role in memory are reviewed elsewhere (Fortress and Frick, 2014). Histone acetylation, phosphorylation, and methylation, along with DNA methylation, are summarized briefly below.

Histone modifications

Histone modifications can alter accessibility of DNA to the transcriptional machinery required for regulating the expression of genes, including those relevant to neuroplasticity and cognition. Histones are primarily modified at the N-terminal portion of their tails that extends beyond the nucleosome. The histone tail can interact with regulatory proteins, as well as neighboring histones and the DNA itself (Tsankova et al., 2007; Marmorstein, 2001). A number of alterations are possible including methylation, phosphorylation, ubiquitination, SUMOylation, and acetylation. Of these, acetylation is perhaps the most well characterized in the neuroepigenetic literature. Acetylation is carried out by histone acetyltransferase (HAT) enzymes, which facilitate the transfer of an acetyl group from acetyl-CoA to lysine residues present on histone tails. In the majority of cases, acetylation of histone tails yields a transcriptionally active state, conferring open euchromatin that is accessible to transcriptional machinery (Tsankova et al., 2007). Conversely, hypoacetylation generally has the opposite effect, largely because replacement of hydrogen with an acetyl group reduces the net positive charge of the histone, reducing its interaction with the negatively charged DNA backbone and increasing the likelihood of nucleosome displacement (Carozza et al., 2003). This displacement opens up the chromatin, making it readily accessible to transcriptional machinery and regulatory factors that can influence transcription and, thereby, regulate gene expression.

As mentioned earlier, activation of signal transduction pathways can facilitate epigenetic changes such as histone acetylation, which may be a mechanism contributing to the precision and the stability of a memory. One example is the ERK/MAPK signaling pathway, which stimulates CREB binding protein (CBP), a transcriptional coactivator with intrinsic HAT activity (Ait-Si-Ali et al., 1999). Evidence that histone acetylation can influence learning was first demonstrated in rodents using the conditioned taste aversion paradigm. Using this task, mice that developed a conditioned taste aversion had increased ERK/MAPK activation and histone acetyltransferase activity in the insular cortex 48 h later (Swank and Sweatt, 2001). Further evidence for the role of histone acetylation in memory was determined in mice with reduced levels of CBP (*cbp*^{+/-} mice), which exhibit impaired long-lasting-LTP, as well as deficits in hippocampus-dependent object recognition memory and contextual fear conditioning (Alarcón et al., 2004). Similar behavioral results were obtained in mice with an inducible dominant-negative system to turn off intrinsic CBP-mediated HAT activity. Interestingly, a histone deacetylase inhibitor rescued memory deficits in these mice, suggesting that reduced histone acetylation could underlie memory deficits in numerous disorders (Korzus et al., 2004). Indeed, mice overexpressing the histone deacetylase HDAC2, which removes acetyl groups from histones, have poor hippocampus-dependent spatial memory and reduced

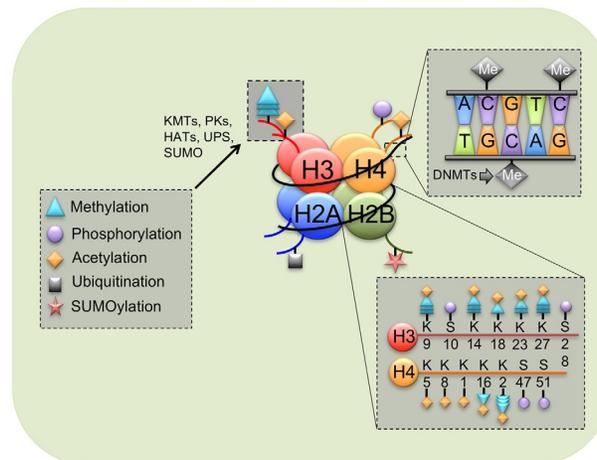


Fig. 3 Diagrammatic illustration of the nucleosome and common epigenetic alterations. The histone octamer consists of two each of histones H2A, H2B, H3, and H4. Abbreviations: *K*, lysine; *S*, serine; *KMTs*, lysine methyltransferases; *PKs*, protein kinases; *HATs*, histone acetyltransferases; *UPS*, ubiquitin proteasome system; *SUMO*, small ubiquitin-like modifier; *Me*, methyl group; *DNMTs*, DNA methyltransferases. Adapted from Fortress, A. M. and Frick, K. M. (2014). Epigenetic regulation of estrogen-dependent memory, *Frontiers in Neuroendocrinology*, **35**(4), 530–549.

synaptic plasticity (Guan et al., 2009). The behavioral deficits induced by overexpressing HDAC2 were reversed by administering an HDAC2 inhibitor, suggesting that memory disorders accompanied by elevated HDAC2 levels could be treated by HDAC2 inhibitors (Guan et al., 2009). Cellular mechanisms responsible for histone acetylation in the hippocampus were determined using contextual fear conditioning in rats. In a series of studies, it was demonstrated that ERK/MAPK signaling, acting upstream of mitogen and stress-activated protein kinase 1 (MSK1), is necessary for the acetylation of H3 (Levenson et al., 2004; Chwang et al., 2007). This evidence collectively suggests that rapid cell-signaling events can lead to changes in epigenetic modifications, such as histone acetylation, to regulate gene transcription and memory.

In addition to histone acetylation, histone phosphorylation is another posttranslational modification poised to regulate memory formation. Protein Serine/Threonine phosphatase 1 (PP1), which dephosphorylates proteins, has been implicated in hippocampal memory impairment and reduced plasticity in rodents (Genoux et al., 2002). In general, phosphorylation is associated with the activation of proteins responsible for a sequence of events that promotes memory and, therefore, interfering with phosphorylation processes could disrupt memory formation. In support of this notion, mice with inducible deficiency of forebrain PP1 demonstrated enhanced long-term memory in the object recognition, object placement, and Morris water maze tasks. This enhanced memory performance coincided with an increase in total phosphorylation of histone H3 on Serine 10 (phospho-H3S10) in addition to increased phospho-H3S10 at the CREB promoter (Koshibu et al., 2009). In addition to its role in regulating histone acetylation, ERK/MAPK signaling can also mediate histone phosphorylation. Activation of the upstream ERK/MAPK regulators PKC and PKA, as well as contextual fear conditioning training, all increased phospho-H3S10 in the hippocampus of rats (Chwang et al., 2006). Further, inhibition of ERK/MAPK prevented phosphorylation of H3S10, suggesting that ERK/MAPK signaling is necessary for H3S10 phosphorylation and contextual fear memory consolidation (Chwang et al., 2006). Notably, H3S10 phosphorylation is also increased in the hippocampus following both retrieval of a fear memory and the phosphorylation of ERK/MAPK (Besnard et al., 2014). Although site-specific dephosphorylation on histone tails has not been demonstrated as a cause of memory impairment, these studies collectively suggest that histone phosphorylation contributes to memory formation.

Whereas histone acetylation and histone phosphorylation are associated with a permissive transcriptional state, histone methylation can be transcriptionally permissive or repressive depending on the residue and number of sites affected. The most commonly examined residue is lysine, which can be mono-(me), di-(me₂), or tri-methylated (me₃) (Ng et al., 2009). Methyl groups are added to residues by histone (lysine) methyltransferases, which are specific to the residue they modify (Black et al., 2012) and are removed by lysine demethylases. For example, unmethylated lysine⁹ on H3 (H3K9) is converted to H3K9me₁ and H3K9me₂ by the G9a methyltransferase, but only the MLL methyltransferase can convert H3K4 to H3K4me₃ (Jarome and Lubin, 2013). H3K4me₃ and H3K9me₂, associated with transcriptional activation and repression, respectively, are both increased in the hippocampus following contextual fear conditioning (Gupta et al., 2010). In support of the role of H3K9me₂ in memory formation, inhibition of the G9a methyltransferase with BIX01294 impairs contextual fear memory formation (Gupta-Agarwal et al., 2012). The importance of the H3K4 MLL methyltransferase in memory has been demonstrated in two independent studies in which mice deficient in *Mll1* exhibited selective impairments in hippocampus-dependent contextual fear memory (Gupta et al., 2010), and mice deficient in *Mll2* displayed impaired hippocampus-dependent object recognition and object placement memory (Kerimoglu et al., 2013). Although these studies suggest that methylation can affect histones in a site-specific manner, future studies will need to investigate the extent to which demethylation is site-specific.

DNA methylation

DNA methylation occurs when a methyl group is added to cytosine residues predominantly within cytosine-guanine (CpG, where 'p' refers to the phosphodiester bond) dinucleotides by a methyl donor (e.g., S-adenosyl methionine; SAM). CpG dense areas are referred to as CpG islands, (Mastroeni et al., 2011). DNA methylation typically represses transcriptional activity when it occurs within the promoter region of a gene, although this is not always the case (Chahrouh et al., 2008). The addition of methyl groups results in the recruitment of corepressor complexes to the DNA, inducing a repressive state by conferring a state of tightly packed heterochromatin that prevents binding of transcription factors and molecules necessary for the initiation of transcription (Tsankova et al., 2007). This often includes the recruitment of histone deacetylases (HDACs), which can act concurrently to affect the acetylation state of neighboring histones, further compacting the chromatin and reducing transcriptional activity. The addition of methyl groups occurs with the aid of DNA methyltransferases or DNMTs (Fig. 3), of which at least two types have been identified. Maintenance methylation, which preserves DNA methylation patterns throughout cellular replication, is catalyzed by DNMT1 (Chouliaras et al., 2010). De novo methyltransferases DNMT3a and DNMT3b are responsible for methylation throughout development regardless of previous methylation status (Okano et al., 1999). DNMT2 is not involved in DNA methylation, but instead plays a role in the methylation of aspartic acid transfer RNA (Chouliaras et al., 2010). Together, these enzymes influence transcription and the expression of a number of genes that are critical for neuroplasticity and memory formation (Miller and Sweatt, 2007; Day et al., 2013a). The dysregulation of these enzymes can lead to detrimental changes in gene expression, which may impair neuronal function and contribute to any number of neuropsychiatric disorders.

Early demonstration for the role of DNA methylation in memory came from evidence showing that contextual fear conditioning increased *Dnmt3a* and *Dnmt3b* mRNA in the hippocampus of rats 30 min after training, and that infusion of a DNMT inhibitor into the hippocampus impaired consolidation of contextual fear memory (Miller and Sweatt, 2007). Interestingly, the increase in methylation was at least partially due to hypermethylation of the memory suppressing *PP1* gene (resulting in a net decrease in expression of *PP1*). The persistence of contextual fear memories were later determined to be maintained through DNA methylation in the dorsomedial prefrontal cortex, a brain region important for long-term memory storage (Miller et al., 2010). Additional

evidence for the role of DNA methylation in regulating memory consolidation came from work linking contextual fear memory with expression of brain derived neurotrophic factor (BDNF), a neurotrophin well known to promote synaptic plasticity and memory (Cowansage et al., 2010). The *Bdnf* gene is unique in that it has multiple exons that can be independently and differentially regulated. Contextual fear conditioning increased the mRNA expression of exon IV, which was associated with hypomethylation at CpG islands corresponding to exon IV (Lubin et al., 2008). Similarly, treatment with a DNMT inhibitor was sufficient to impair memory, increase methylation of the *Bdnf* gene, and decrease *Bdnf* gene transcription (Lubin et al., 2008). These findings support the conclusion that manipulation of DNA methylation is yet another method of regulating memory.

Estrogenic regulation of histone modifications and effects on memory

As mentioned earlier in this chapter, the ability of a dorsal hippocampal E_2 infusion to enhance hippocampal memory consolidation depends on the rapid activation of numerous cell-signaling cascades, including ERK/MAPK, in the hippocampus (Fernandez et al., 2008; Fortress et al., 2013). Activation of ERK in the hippocampus leads to an increase in the acetylation and phosphorylation of the histone protein H3, and these increases are associated with memory formation (Levenson et al., 2004; Chwang et al., 2006). Thus, our laboratory reasoned that E_2 might enhance memory by triggering ERK-mediated histone alterations. Indeed, direct infusion of E_2 into the DH increases HAT activity in the DH within 15 min and increases DH H3K9,14 acetylation within 30 min (Zhao et al., 2010, 2012). However, E_2 infusion into the DH does not affect acetylation of H2BK12 or H4K12 (Zhao et al., 2010, 2012), suggesting an effect specific to H3K9,14. Additionally, DH infusion of an ERK inhibitor impaired memory and blocked the E_2 -induced increase in H3K9,14 acetylation (Zhao et al., 2010), suggesting that H3 acetylation is critical for estrogenic regulation of hippocampal memory consolidation. To determine the extent to which histone acetylation is necessary for E_2 to enhance memory, the HAT inhibitor garcinol was coinjected with E_2 . Garcinol prevented DH-infused E_2 from elevating H3K9,14 acetylation and enhancing object memory consolidation (Zhao et al., 2012), demonstrating that H3 acetylation is necessary for the estrogenic enhancement of object memory formation.

E_2 may regulate histone acetylation in the long term by altering the expression of HDAC proteins. Four hours after DH infusion, E_2 significantly decreases levels of HDAC2 and HDAC3 protein (Zhao et al., 2012; Fortress et al., 2014), both of which have been implicated as negative regulators of hippocampus-dependent memory (Guan et al., 2009; McQuown et al., 2011; Graff et al., 2012). E_2 -induced downregulation of HDAC2 protein expression was blocked by garcinol (Zhao et al., 2012), suggesting that HAT activity regulates expression of this memory repressing HDAC. Collectively, these findings suggest that E_2 may exert its beneficial effects on memory by increasing H3 acetylation and reducing the expression of negative regulators of memory such as HDAC2 and HDAC3 (Fig. 2).

Many of the specific gene targets affected by E_2 -mediated H3 acetylation remain to be defined, although genes involved in neuroplasticity are likely candidates. One such example is the neurotrophin BDNF, which has an established role in regulating spine density, synaptic plasticity, and memory (Scharfman et al., 2003; Heldt et al., 2007; Spencer et al., 2008; Luine and Frankfurt, 2013). Recent evidence demonstrates that E_2 infusion directly into the DH specifically increases H3 acetylation at *Bdnf* promoters pII and pIV 30 min after infusion in both young and middle-aged ovariectomized mice (Fortress et al., 2014). These alterations in H3 acetylation at pII and pIV precede a significant increase in BDNF and pro-BDNF protein levels that occurs in the DH 4 and 6 h after infusion. Thus, one potential mechanism through which E_2 exerts its beneficial mnemonic effects is via epigenetic regulation of BDNF in the hippocampus. However, many genes are involved in memory formation, so additional research is needed to elucidate how histone alterations may impact the expression of other genes that contribute to the beneficial effects of E_2 on memory.

Estrogenic regulation of DNA methylation and effects on memory

DNA methylation can also be modified by E_2 treatment. For example, infusion of E_2 directly into the DH significantly increases *Dnmt3a* and *Dnmt3b* mRNA levels 45 min later, however, only DNMT3b protein levels were significantly elevated 4 h later (Zhao et al., 2010). Protein and mRNA levels of the maintenance enzyme DNMT1 remained unaffected after direct DH infusion of E_2 . Interestingly, E_2 -mediated increases in DNMT3b protein were blocked by coadministration of the HAT inhibitor garcinol (Zhao et al., 2012), suggesting that histone acetylation is required for E_2 -mediated changes in DNMT3b protein. This finding is consistent with other research demonstrating that interactions between DNA methylation and histone acetylation are critical for modulating cognition and neuroplasticity (Miller et al., 2008). The E_2 -mediated enhancement of object memory consolidation is also blocked by a DH infusion of the DNMT inhibitor 5-aza-2'-deoxycytidine (Zhao et al., 2010), suggesting that DNA methylation is also critical for the memory-enhancing effects of E_2 (Fig. 2). However, the specific genes and CpG sites methylated by E_2 in the DH remain unknown and this is an area ripe for future research.

Gaps in Knowledge and Implications for Future Studies

Understanding the mechanisms through which E_2 can orchestrate epigenetic processes to regulate memory is important for understanding cognitive function in both health and disease. Thus far, these mechanisms have only begun to be elucidated as outlined in this chapter. The extent to which E_2 regulates other posttranslational modifications such as histone phosphorylation, ubiquitination, methylation, or SUMOylation in the hippocampus through various cell-signaling mechanisms remains to be identified. Further, only very few gene targets important for memory whose expression is epigenetically regulated by E_2 have been identified to date. Thus, understanding which epigenetic modifications are regulated by E_2 and the downstream transcriptional

and translational consequences of these modifications will be critical next steps. Finally, understanding the functional implications of estrogenic regulation of neuroepigenetics will be imperative to better understanding the etiology of disease and improving the design of drugs for reducing memory dysfunction in the future.

It's also important to consider that it is very unlikely that the epigenetic effects of E₂ on memory are restricted to the hippocampus. Parallel evidence for estrogenic regulation of epigenetic processes exists in other brain regions outside of the hippocampus, including the amygdala and the prefrontal cortex. For example, both the prefrontal cortex and the amygdala are actively being investigated for sensitivity to epigenetic modifications in neuropsychiatric disorders, including PTSD (Pizzimenti and Lattal, 2015). E₂ facilitates extinction learning following fear conditioning, which corresponds to decreased activation in the amygdala and increased activation in the ventromedial prefrontal cortex in rats (Zeidan et al., 2011). Although a necessary role for E₂ in regulating epigenetic mechanisms in the fear memory circuitry has not been tested, a necessary role of histone acetylation in fear memory consolidation was demonstrated using direct infusion of the HAT inhibitor garcinol into the lateral amygdala, which impaired fear memory consolidation in rats (Maddox et al., 2013). Mice that were in the metestrus phase of the estrous cycle (low estrogen state) or were ovariectomized exhibited increased *Hdac4* mRNA in the amygdala following cued fear conditioning, suggesting a potential mechanism through which low levels of E₂ could impair processing of cues related to PTSD (Maddox et al., 2017). These findings suggest that E₂ may promote fear memory consolidation by increasing histone acetylation in the other brain areas, however direct evidence has not been provided.

Finally, the role of de novo neurosteroidogenesis in regulating the epigenetic processes that govern memory formation remains unknown and is an area of increasing clinical relevance. All sex steroid hormones, including E₂, are synthesized in the brain, including the hippocampus, and suppressing endogenous E₂ synthesis with aromatase inhibitors prevents memory formation in female mice and male song birds (Bailey et al., 2013; Tuscher et al., 2016b). Similarly, clinical studies suggest that aromatase inhibition may negatively impact cognitive function in human females. Aromatase inhibitors such as letrozole are used to treat hormone-receptor positive forms of breast cancer (Geisler et al., 2002; Puddefoot et al., 2002), and some findings suggest that such treatments compromise working memory, concentration, and performance in verbal and visual memory tasks (Collins et al., 2009; Bender et al., 2007, 2015). Epigenetic regulation of the aromatase enzyme and downstream epigenetic changes in E₂-mediated memory in response to aromatase inhibitors have yet to be identified, but may prove to be important given increasing evidence that aromatase inhibition leads to memory impairment in females.

Summary

The past 30 years has provided exciting new information about the molecular mechanisms underlying memory formation and dysfunction. In particular, neuroepigenetic studies have illuminated the complexities of gene regulation and revealed a multitude of ways in which epigenetic processes can alter behavior without changing the genetic code. Combined with behavioral neuroendocrinology, neuroepigenetics is advancing our understanding of how hormones regulate the epigenetic processes that influence behavior. Such regulation could explain how environmental, chromosomal, and psychological factors determine individual responses to specific situations. Although information on E₂-mediated epigenetic alterations in the brain remains limited, there is ample potential to explore how sex-steroid hormones modulate behavior and disease in numerous brain regions with implications across a spectrum of disorders. It is our hope that this is where the future will take us and other investigators.

References

- Acosta JI, Mayer L, Talboom JS, Tsang CW, Smith CJ, Enders CK, and Bimonte-Nelson HA (2009) Transitional versus surgical menopause in a rodent model: Etiology of ovarian hormone loss impacts memory and the acetylcholine system. *Endocrinology* 150(9): 4248–4259.
- Acosta JI, Mayer LP, Braden BB, Nonnenmacher S, Mennenga SE, and Bimonte-Nelson HA (2010) The cognitive effects of conjugated equine estrogens depend on whether menopause etiology is transitional or surgical. *Endocrinology* 151(8): 3795–3804.
- Adams JP and Sweatt JD (2002) Molecular psychology: Roles for the ERK MAP kinase cascade in memory. *Annual Review of Pharmacology and Toxicology* 42: 135–163.
- Ait-Si-Ali S, Carlisi D, Ramirez S, Upegui-Gonzalez LC, Duquet A, Robin P, Rudkin B, Harel-Bellan A, and Trouche D (1999) Phosphorylation by p44 MAP kinase/ERK1 stimulates CBP histone acetyl transferase activity in vitro. *Biochemical Biophysical Research Communications* 262(1): 157–162.
- Alarcón JM, Malleret G, Touzani K, Vronskaya S, Ishii S, Kandel ER, and Barco A (2004) Chromatin acetylation, memory, and LTP are impaired in CBP +/- mice: A model for the cognitive deficit in Rubinstein-Taybi syndrome and its amelioration. *Neuron* 42: 947–959.
- Albert K, Pruessner J, and Newhouse P (2015) Estradiol levels modulate brain activity and negative responses to psychosocial stress across the menstrual cycle. *Psychoneuroendocrinology* 59: 14–24.
- Asl SZ, Khaksari M, Khachki AS, Shahrokhi N, and Nourzade S (2013) Contribution of estrogen receptors alpha and beta in the brain response to traumatic brain injury. *Journal of Neurosurgery* 119(2): 353–361.
- Bailey DJ, Ma C, Soma KK, and Saldanha CJ (2013) Inhibition of hippocampal aromatization impairs spatial memory performance in a male songbird. *Endocrinology* 154: 4707–4714.
- Barha CK, Dalton GL, and Galea LA (2010) Low doses of 17alpha-estradiol and 17beta-estradiol facilitate, whereas higher doses of estrone and 17alpha- and 17beta-estradiol impair, contextual fear conditioning in adult female rats. *Neuropsychopharmacology* 35(2): 547–559.
- Becker JB (2016) Sex differences in addiction. *Dialogues in Clinical Neuroscience* 18(4): 395–402.
- Bender CM, Sereika SM, Brufsky AM, Ryan CM, Vogel VG, Rastogi P, Cohen SM, Casillo FE, and Berga SL (2007) Memory impairments with adjuvant anastrozole versus tamoxifen in women with early-stage breast cancer. *Menopause* 14(6): 995–998.
- Bender CM, Merriman JD, Gentry AL, Ahrendt GM, Berga SL, Brufsky AM, Casillo FE, Dailey MM, Erickson KI, Kratochvil FM, McAuliffe PF, Rosenzweig MQ, Ryan CM, and Sereika SM (2015) Patterns of change in cognitive function with anastrozole therapy. *Cancer* 121(15): 2627–2636.

- Besnard A, Laroche S, and Caboche J (2014) Comparative dynamics of MAPK/ERK signalling components and immediate early genes in the hippocampus and amygdala following contextual fear conditioning and retrieval. *Brain Structure and Function* 219(1): 415–430.
- Bimonte HA, Granholm A-CE, Seo H, and Isacson O (2002) Spatial memory testing decreases hippocampal amyloid precursor protein in young, but not aged, female rats. *Neuroscience Letters* 298: 50–54.
- Bimonte-Nelson HA, Acosta JI, and Talboom JS (2010) Neuroscientists as cartographers: Mapping the crossroads of gonadal hormones, memory and age using animal models. *Molecules* 15(9): 6050–6105.
- Black JC, Van Rechem C, and Whetstone JR (2012) Histone lysine methylation dynamics: Establishment, regulation, and biological impact. *Molecular Cell* 48(4): 491–507.
- Boulware MI, Weick JP, Becklund BR, Kuo SP, Groth RD, and Mermelstein PG (2005) Estradiol activates group I and II metabotropic glutamate receptor signaling, leading to opposing influences on cAMP response element-binding protein. *Journal of Neuroscience* 25: 5066–5078.
- Boulware MI, Heisler JD, and Frick KM (2013) The memory-enhancing effects of hippocampal estrogen receptor activation involve metabotropic glutamate receptor signaling. *Journal of Neuroscience* 33(38): 15184–15194.
- Bowman RE, Ferguson D, and Luine VN (2002) Effects of chronic restraint stress and estradiol on open field activity, spatial memory, and monoaminergic neurotransmitters in ovariectomized rats. *Neuroscience* 113(2): 401–410.
- Carrozza MJ, Utley RT, Workman JL, and Cote J (2003) The diverse functions of histone acetyltransferase complexes. *Trends in Genetics* 19(6): 321–329.
- Chahrour M, Jung SY, Shaw C, Zhou X, Wong ST, Qin J, and Zoghbi HY (2008) MeCP2, a key contributor to neurological disease, activates and represses transcription. *Science* 320(5880): 1224–1229.
- Chang YJ, Yang CH, Liang YC, Yeh CM, Huang CC, and Hsu KS (2009) Estrogen modulates sexually dimorphic contextual fear extinction in rats through estrogen receptor beta. *Hippocampus* 19(11): 1142–1150.
- Chouliaras L, Rutten BP, Kenis G, Peerbooms O, Visser PJ, Verhey F, van Os J, Steinbusch HW, and van den Hove DL (2010) Epigenetic regulation in the pathophysiology of Alzheimer's disease. *Progress in Neurobiology* 90(4): 498–510.
- Chwang WB, O'Riordan KJ, Levenson JM, and Sweatt JD (2006) ERK/MAPK regulates hippocampal histone phosphorylation following contextual fear conditioning. *Learning and Memory* 13(3): 322–328.
- Chwang WB, Arthur JS, Schumacher A, and Sweatt JD (2007) The nuclear kinase mitogen- and stress-activated protein kinase 1 regulates hippocampal chromatin remodeling in memory formation. *Journal of Neuroscience* 27(46): 12732–12742.
- Collins B, Mackenzie J, Stewart A, Bielajew C, and Verma S (2009) Cognitive effects of hormonal therapy in early stage breast cancer patients: A prospective study. *Psychooncology* 18(8): 811–821.
- Cowsavage KK, LeDoux JE, and Monfils MH (2010) Brain-derived neurotrophic factor: A dynamic gatekeeper of neural plasticity. *Current Molecular Pharmacology* 3(1): 12–29.
- Crick F (1984) Memory and molecular turnover. *Nature* 312(5990): 101.
- Daniel JM (2006) Effects of oestrogen on cognition: What have we learned from basic research? *Journal of Neuroendocrinology* 18: 787–795.
- Daniel JM, Fader AJ, Spencer AL, and Dohanich GP (1997) Estrogen enhances performance of female rats during acquisition of a radial arm maze. *Hormones and Behavior* 32: 217–225.
- Daniel JM, Roberts SL, and Dohanich GP (1999) Effects of ovarian hormones and environment on radial maze and water maze performance of female rats. *Physiology and Behavior* 66: 11–20.
- Day JJ, Childs D, Guzman-Karlsson MC, Kibe M, Moulden J, Song E, Tahir A, and Sweatt JD (2013a) DNA methylation regulates associative reward learning. *Nature Neuroscience* 16(10): 1445–1452.
- Day NL, Floyd CL, D'Alessandro TL, Hubbard WJ, and Chaudry IH (2013b) 17beta-estradiol confers protection after traumatic brain injury in the rat and involves activation of G protein-coupled estrogen receptor 1. *Journal of Neurotrauma* 30(17): 1531–1541.
- Dye RV, Miller KJ, Singer EJ, and Levine EJ (2012) Hormone replacement therapy and risk for neurodegenerative diseases. *International Journal of Alzheimer's Disease* 258454.
- Fader AJ, Hendricson AW, and Dohanich GP (1998) Estrogen improves performance of reinforced T-maze alternation and prevents the amnesic effects of scopolamine administered systemically or intrahippocampally. *Neurobiology of Learning and Memory* 69: 225–240.
- Fader AJ, Johnson PEM, and Dohanich GP (1999) Estrogen improves working but not reference memory and prevents amnesic effects of scopolamine on a radial-arm maze. *Pharmacology Biochemistry and Behavior* 62: 711–717.
- Fan L, Zhao Z, Orr PT, Chambers CH, Lewis MC, and Frick KM (2010) Estradiol-induced object memory consolidation in middle-aged female mice requires dorsal hippocampal extracellular signal-regulated kinase and phosphatidylinositol 3-kinase activation. *Journal of Neuroscience* 30: 4390–4400.
- Fernandez SM, Lewis MC, Pechenino AS, Harburger LL, Orr PT, Gresack JE, Schafe GE, and Frick KM (2008) Estradiol-induced enhancement of object memory consolidation involves hippocampal ERK activation and membrane-bound estrogen receptors. *Journal of Neuroscience* 28: 8660–8667.
- Fortress AM and Frick KM (2014) Epigenetic regulation of estrogen-dependent memory. *Frontiers in Neuroendocrinology* 35(4): 530–549.
- Fortress AM, Fan L, Orr PT, Zhao Z, and Frick KM (2013) Estradiol-induced object recognition memory consolidation is dependent on activation of mTOR signaling in dorsal hippocampus. *Learning and Memory* 20(3): 147–155.
- Fortress AM, Kim J, Poole RL, Gould TJ, and Frick KM (2014) 17β-estradiol regulates histone alterations associated with memory consolidation and increases Bdnf promoter acetylation in middle-aged female mice. *Learning and Memory* 21(9): 457–467.
- Fox M, Berzuini C, and Knapp LA (2013) Cumulative estrogen exposure, number of menstrual cycles, and Alzheimer's risk in a cohort of British women. *Psychoneuroendocrinology* 38(12): 2973–2982.
- Frick KM (2009) Estrogens and age-related memory decline in rodents: What have we learned and where do we go from here? *Hormones and Behavior* 55(1): 2–23.
- Frick KM (2012) Building a better hormone therapy? How understanding the rapid effects of sex steroid hormones could lead to new therapeutics for age-related memory decline. *Behavioral Neuroscience* 126(1): 29–53.
- Geisler J, Haynes B, Anker G, Dowsett M, and Lonning PE (2002) Influence of letrozole and anastrozole on total body aromatization and plasma estrogen levels in postmenopausal breast cancer patients evaluated in a randomized, cross-over study. *Journal of Clinical Oncology* 20(3): 751–757.
- Genoux D, Haditsch U, Knobloch M, Michalon A, Storm D, and Mansuy IM (2002) Protein phosphatase 1 is a molecular constraint on learning and memory. *Nature* 418(6901): 970–975.
- Gillies GE and McArthur S (2010) Estrogen actions in the brain and the basis for differential action in men and women: A case for sex-specific medicines. *Pharmacological Reviews* 62(2): 155–198.
- Gould E, Westlind-Danielsson A, Frankfurt M, and McEwen BS (1990) Sex differences and thyroid hormone sensitivity of hippocampal pyramidal cells. *Journal of Neuroscience* 10: 996–1003.
- Graff J, Rei D, Guan JS, Wang WY, Seo J, Hennig KM, Nieland TJ, Fass DM, Kao PF, Kahn M, Su SC, Samiei A, Joseph N, Haggarty SJ, Delalle I, and Tsai LH (2012) An epigenetic blockade of cognitive functions in the neurodegenerating brain. *Nature* 483(7388): 222–226.
- Guan JS, Haggarty SJ, Giacometti E, Dannenberg JH, Joseph N, Gao J, Nieland TJ, Zhou Y, Wang X, Mazitschek R, Bradner JE, DePinho RA, Jaenisch R, and Tsai LH (2009) HDAC2 negatively regulates memory formation and synaptic plasticity. *Nature* 459: 55–60.
- Gupta S, Kim SY, Artis S, Molfese DL, Schumacher A, Sweatt JD, Paylor RE, and Lubin FD (2010) Histone methylation regulates memory formation. *Journal of Neuroscience* 30: 3589–3599.
- Gupta-Agarwal S, Franklin AV, Deramus T, Wheelock M, Davis RL, McMahon LL, and Lubin FD (2012) G9a/GLP histone lysine dimethyltransferase complex activity in the hippocampus and the entorhinal cortex is required for gene activation and silencing during memory consolidation. *Journal of Neuroscience* 32(16): 5440–5453.
- Hammond R and Gibbs RB (2011) GPR30 is positioned to mediate estrogen effects on basal forebrain cholinergic neurons and cognitive performance. *Brain Research* 1379: 53–60.

- Hampson E (1990) Estrogen-related variations in human spatial and articulatory-motor skills. *Psychoneuroendocrinology* 15: 97–111.
- Heldring N, Pike A, Andersson S, Matthews J, Cheng G, Hartman J, Tujague M, Strom A, Treuter E, Warner M, and Gustafsson JA (2007) Estrogen receptors: How do they signal and what are their targets. *Physiological Reviews* 87(3): 905–931.
- Heldt SA, Stanek L, Chhatwal JP, and Ressler KJ (2007) Hippocampus-specific deletion of BDNF in adult mice impairs spatial memory and extinction of aversive memories. *Molecular Psychiatry* 12: 656–670.
- Hoefler CA and Klann E (2010) mTOR signaling: At the crossroads of plasticity, memory and disease. *Trends in Neuroscience* 33: 67–75.
- Holliday R (1999) Is there an epigenetic component in long-term memory? *Journal of Theoretical Biology* 200(3): 339–341.
- Horwood JM, Dufour F, Laroche S, and Davis S (2006) Signalling mechanisms mediated by the phosphoinositide 3-kinase/Akt cascade in synaptic plasticity and memory in the rat. *European Journal of Neuroscience* 23: 3375–3384.
- Impey S, Smith DM, Obrietan K, Donahue R, Wade C, and Storm DR (1998) Stimulation of cAMP response element (CRE)-mediated transcription during contextual learning. *Nature Neuroscience* 1: 595–601.
- Jarome TJ and Lubin FD (2013) Histone lysine methylation: Critical regulator of memory and behavior. *Reviews in the Neurosciences*: 1–13.
- Jensen EV (1962) On the mechanism of estrogen action. *Perspectives in Biology and Medicine* 6: 47–59.
- Kerimoglu C, Agis-Balboa RC, Kranz A, Stilling R, Bahari-Javan S, Benito-Garagorri E, Halder R, Burkhardt S, Stewart AF, and Fischer A (2013) Histone-methyltransferase MLL2 (KMT2B) is required for memory formation in mice. *Journal of Neuroscience* 33(8): 3452–3464.
- Korzus E, Rosenfeld MG, and Mayford M (2004) CBP histone acetyltransferase activity is a critical component of memory consolidation. *Neuron* 42: 961–972.
- Koshibu K, Graff J, Beullens M, Heitz FD, Berchtold D, Russig H, Farinelli M, Bollen M, and Mansuy IM (2009) Protein phosphatase 1 regulates the histone code for long-term memory. *Journal of Neuroscience* 29(41): 13079–13089.
- Laurer LJ, Andersen K, Dewey ME, Letenneur L, Ott A, Amaducci LA, Brayne C, Copeland JRM, Dartigues J-F, Kragh-Sorensen P, Lobo A, Martinez-Lage JM, Stijnen T, and Hofman A (1999) Rates and risk factors for dementia and Alzheimer's disease. *Neurology* 52: 78–84.
- Lebron-Milad K and Milad MR (2012) Sex differences, gonadal hormones and the fear extinction network: Implications for anxiety disorders. *Biology of Mood and Anxiety Disorders* 2(1): 3.
- Leuner B, Mendolia-Loffredo S, and Shors TJ (2004) High levels of estrogen enhance associative memory formation in ovariectomized females. *Psychoneuroendocrinology* 29: 883–890.
- Levenson JM, O'Riordan KJ, Brown KD, Trinh MA, Molfese DL, and Sweatt JD (2004) Regulation of histone acetylation during memory formation in the hippocampus. *Journal of Biological Chemistry* 279(39): 40545–40559.
- Lewis MC, Kerr KM, Orr PT, and Frick KM (2008) Estradiol-induced enhancement of object memory consolidation involves NMDA receptors and protein kinase A in the dorsal hippocampus of female C57BL/6 mice. *Behavioral Neuroscience* 122: 716–721.
- Loy R, Gerlach JL, and McEwen BS (1988) Autoradiographic localization of oestradiol binding neurons in the hippocampal formation and the entorhinal cortex. *Developmental Brain Research* 39: 245–251.
- Lubin FD, Roth TL, and Sweatt JD (2008) Epigenetic regulation of BDNF gene transcription in the consolidation of fear memory. *Journal of Neuroscience* 28(42): 10576–10586.
- Luine VN (2014) Estradiol and cognitive function: Past, present and future. *Hormones and Behavior* 66(4): 602–618.
- Luine V and Frankfurt M (2013) Interactions between estradiol, BDNF and dendritic spines in promoting memory. *Neuroscience* 239: 34–45.
- Luine VN, Richards ST, Wu VY, and Beck KD (1998) Estradiol enhances learning and memory in a spatial memory task and effects levels of monoaminergic neurotransmitters. *Hormones and Behavior* 34(2): 149–162.
- Luine VN, Jacome LF, and Macluskus NJ (2003) Rapid enhancement of visual and place memory by estrogens in rats. *Endocrinology* 144(7): 2836–2844.
- Maddox SA, Watts CS, Doyere V, and Schafe GE (2013) A naturally-occurring histone acetyltransferase inhibitor derived from *Garcinia indica* impairs newly acquired and reactivated fear memories. *PLoS One* 8(1): e54463.
- Maddox SA, Kilaru V, Shin J, Jovanovic T, Almi LM, Dias BG, Norrholm SD, Fani N, Michopoulos V, Ding Z, Conneely KN, Binder EB, Ressler KJ, and Smith AK (2017) Estrogen-dependent association of HDAC4 with fear in female mice and women with PTSD. *Molecular Psychiatry* 23(3): 658–665.
- Maggi A, Susanna L, Bettini E, Mantero G, and Zucchi I (1989) Hippocampus: A target for estrogen action in mammalian brain. *Molecular Endocrinology* 3(7): 1165–1170.
- Maillet D and Rajah MN (2013) Association between prefrontal activity and volume change in prefrontal and medial temporal lobes in aging and dementia: A review. *Ageing Research Reviews* 12(2): 479–489.
- Maki PM (2012) Minireview: Effects of different HT formulations on cognition. *Endocrinology* 153(8): 3564–3570.
- Maki PM, Rich JB, and Rosenbaum RS (2002) Implicit memory varies across the menstrual cycle: Estrogen effects in young women. *Neuropsychologia* 40(5): 518–529.
- Marmorstein R (2001) Structure of histone acetyltransferases. *Journal of Molecular Biology* 311(3): 433–444.
- Mastroeni D, Grover A, Delvaux E, Whiteside C, Coleman PD, and Rogers J (2011) Epigenetic mechanisms in Alzheimer's disease. *Neurobiology of Aging* 32(7): 1161–1180.
- Mazzeo EA and Soliman KF (2012) Basic concepts of epigenetics: Impact of environmental signals on gene expression. *Epigenetics* 7(2): 119–130.
- McQuown SC, Barrett RM, Matheos DP, Post RJ, Rogge GA, Alenghat T, Mullican SE, Jones S, Rusche JR, Lazar MA, and Wood MA (2011) HDAC3 is a critical negative regulator of long-term memory formation. *Journal of Neuroscience* 31(2): 764–774.
- Milad MR, Zeidan MA, Contero A, Pitman RK, Klibanski A, Rauch SL, and Goldstein JM (2010) The influence of gonadal hormones on conditioned fear extinction in healthy humans. *Neuroscience* 168(3): 652–658.
- Miller CA and Sweatt JD (2007) Covalent modification of DNA regulates memory formation. *Neuron* 53: 857–869.
- Miller CA, Campbell SL, and Sweatt JD (2008) DNA methylation and histone acetylation work in concert to regulate memory formation and synaptic plasticity. *Neurobiology of Learning and Memory* 89: 599–603.
- Miller CA, Gavin CF, White JA, Parrish RR, Honasoge A, Yancey CR, Rivera IM, Rubio MD, Rumbaugh G, and Sweatt JD (2010) Cortical DNA methylation maintains remote memory. *Nature Neuroscience* 13(6): 664–666.
- Nelson RJ (2000) *An Introduction to Behavioral Endocrinology*. Sunderland, MA: Sinauer Associates.
- Ng SS, Yue WW, Oppermann U, and Klose RJ (2009) Dynamic protein methylation in chromatin biology. *Cellular and Molecular Life Sciences* 66(3): 407–422.
- Okano M, Bell DW, Haber DA, and Li E (1999) DNA methyltransferases Dnmt3a and Dnmt3b are essential for de novo methylation and mammalian development. *Cell* 99(3): 247–257.
- Packard MG and Teather LA (1997) Intra-hippocampal estradiol infusion enhances memory in ovariectomized rats. *Neuroreport* 8: 3009–3013.
- Phan A, Gabor CS, Favaro KJ, Kaschack S, Armstrong JN, MacLuskus NJ, and Choleris E (2012) Low doses of 17beta-estradiol rapidly improve learning and increase hippocampal dendritic spines. *Neuropsychopharmacology* 37(10): 2299–2309.
- Phillips SM and Sherwin BB (1992) Variations in memory function and sex steroid hormones across the menstrual cycle. *Psychoneuroendocrinology* 17(5): 497–506.
- Pizzimenti CL and Lattal KM (2015) Epigenetics and memory: Causes, consequences and treatments for post-traumatic stress disorder. *Genes, Brain and Behavior* 14(1): 73–84.
- Puddefoot JR, Barker S, Glover HR, Malouite SD, and Vinson GP (2002) Non-competitive steroid inhibition of oestrogen receptor functions. *International Journal of Cancer* 101(1): 17–22.
- Rhodes ME and Frye CA (2004) Estrogen has mnemonic-enhancing effects in the inhibitory avoidance task. *Pharmacology Biochemistry Behavior* 78(3): 551–558.
- Rocca WA, Bower JH, Maraganore DM, Ahlskog JE, Grossardt BR, de Andrade M, and Melton LJ 3rd (2007) Increased risk of cognitive impairment or dementia in women who underwent oophorectomy before menopause. *Neurology* 69(11): 1074–1083.
- Rocca WA, Grossardt BR, and Shuster LT (2011) Oophorectomy, menopause, estrogen treatment, and cognitive aging: Clinical evidence for a window of opportunity. *Brain Research* 1379: 188–198.
- Rothbart SB and Strahl BD (2014) Interpreting the language of histone and DNA modifications. *Biochimica et Biophysica Acta* 1839(8): 627–643.

- Sampath D, Sathyanesan M, and Newton SS (2017) Cognitive dysfunction in major depression and Alzheimer's disease is associated with hippocampal-prefrontal cortex dysconnectivity. *Neuropsychiatric Disease and Treatment* 13: 1509–1519.
- Sandstrom NJ and Williams CL (2004) Spatial memory retention is enhanced by acute and continuous estradiol replacement. *Hormones and Behavior* 45: 128–135.
- Scharfman HE, Mercurio TC, Goodman JH, Wilson MA, and MacLusky NJ (2003) Hippocampal excitability increases during the estrous cycle in the rat: A potential role for brain-derived neurotrophic factor. *Journal of Neuroscience* 23: 11641–11652.
- Schlinger BA and Remage-Healey L (2012) Neurosteroidogenesis: Insights from studies of songbirds. *Journal of Neuroendocrinology* 24(1): 16–21.
- Sherwin BB and Henry JF (2008) Brain aging modulates the neuroprotective effects of estrogen on selective aspects of cognition in women: A critical review. *Frontiers in Neuroendocrinology* 29(1): 88–113.
- Silva AJ, Paylor R, Wehner JM, and Tonegawa S (1992) Impaired spatial learning in α -calcium-calmodulin kinase II mutant mice. *Science* 257: 206–211.
- Singh M, Meyer EM, Millard WJ, and Simpkins JW (1994) Ovarian steroid deprivation results in a reversible learning impairment and compromised cholinergic function in female Sprague-Dawley rats. *Brain Research* 644: 305–312.
- Smith CG and McMahon LL (2005) Estrogen-induced increase in the magnitude of long-term potentiation occurs only when the ratio of NMDA transmission to AMPA transmission is increased. *Journal of Neuroscience* 25(34): 7780–7791.
- Small SA, Schobel SA, Buxton RB, Witter MP, and Barnes CA (2011) A pathophysiological framework of hippocampal dysfunction in ageing and disease. *Nature Reviews Neuroscience* 12(10): 585–601.
- Spencer JL, Waters EM, Romeo RD, Wood GE, Milner TA, and McEwen BS (2008) Uncovering the mechanisms of estrogen effects on hippocampal function. *Frontiers in Neuroendocrinology* 29(2): 219–237.
- Srivastava DP and Evans PD (2013) G-protein oestrogen receptor 1: Trials and tribulations of a membrane oestrogen receptor. *Journal of Neuroendocrinology* 25(11): 1219–1230.
- Swank MW and Sweatt JD (2001) Increased histone acetyltransferase and lysine acetyltransferase activity and biphasic activation of the ERK/RSK cascade in insular cortex during novel taste learning. *Journal of Neuroscience* 21(10): 3383–3391.
- Tsankova N, Renthal W, Kumar A, and Nestler EJ (2007) Epigenetic regulation in psychiatric disorders. *Nature Reviews Neuroscience* 8(5): 355–367.
- Tuscher JJ, Fortress AM, Kim J, and Frick KM (2015) Regulation of novel object recognition and object placement by ovarian sex steroid hormones. *Behavioural Brain Research* 285: 140–157.
- Tuscher JJ, Luine V, Frankfurt M, and Frick KM (2016a) Estradiol-mediated spine changes in the dorsal Hippocampus and medial prefrontal cortex of ovariectomized female mice depend on ERK and mTOR activation in the dorsal hippocampus. *Journal of Neuroscience* 36(5): 1483–1489.
- Tuscher JJ, Szinte JS, Starrett JR, Krentzel AA, Fortress AM, Remage-Healey L, and Frick KM (2016b) Inhibition of local estrogen synthesis in the hippocampus impairs hippocampal memory consolidation in ovariectomized female mice. *Hormones and Behavior* 83: 60–67.
- Walf AA, Koonce CJ, and Frye CA (2008) Estradiol or diarylpropionitrile administration to wild type, but not estrogen receptor beta knockout, mice enhances performance in the object recognition and object placement tasks. *Neurobiology of Learning and Memory* 89: 513–521.
- Woolley CS (2007) Acute effects of estrogen on neuronal physiology. *Annual Review of Pharmacology and Toxicology* 47: 657–680.
- Woolley CS and McEwen BS (1992) Estradiol mediates fluctuation in hippocampal synapse density during the estrous cycle in the adult rat. *Journal of Neuroscience* 12: 2549–2554.
- Woolley CS and McEwen BS (1993) Roles of estradiol and progesterone in regulation of hippocampal dendritic spine density during the estrous cycle in the rat. *Journal of Comparative Neurology* 336: 293–306.
- Woolley CS, Gould E, Frankfurt M, and McEwen BS (1990) Naturally occurring fluctuation in dendritic spine density on adult hippocampal pyramidal neurons. *Journal of Neuroscience* 10: 4035–4039.
- Zeidan MA, Igoe SA, Linnman C, Vitalo A, Levine JB, Klibanski A, Goldstein JM, and Milad MR (2011) Estradiol modulates medial prefrontal cortex and amygdala activity during fear extinction in women and female rats. *Biological Psychiatry* 70(10): 920–927.
- Zhao Z, Fan L, and Frick KM (2010) Epigenetic alterations regulate the estradiol-induced enhancement of memory consolidation. *Proceedings of the National Academy of Sciences USA* 107: 5605–5610.
- Zhao Z, Fan L, Fortress AM, Boulware MI, and Frick KM (2012) Hippocampal histone acetylation regulates object recognition and the estradiol-induced enhancement of object recognition. *Journal of Neuroscience* 32(7): 2344–2351.

Further Reading

- Colciago A and Magnaghi V (2016) Neurosteroids involvement in the epigenetic control of memory formation and storage. *Neural Plasticity* 2016: 5985021.
- Cover KK, Maeng LY, Lebron-Milad K, and Milad MR (2014) Mechanisms of estradiol in fear circuitry: Implications for sex differences in psychopathology. *Translational Psychiatry* 4: e422.
- Daniel JM, Hulst JL, and Berbling JL (2006) Estradiol replacement enhances working memory in middle-aged rats when initiated immediately after ovariectomy but not after a long-term period of ovarian hormone deprivation. *Endocrinology* 147: 607–614.
- Hawkins RD and Byrne JH (2015) Associative learning in invertebrates. *Cold Spring Harbor Perspectives in Biology* 7(5).
- McCarthy MM and Nugent BM (2015) At the frontier of epigenetics of brain sex differences. *Frontiers in Behavioral Neuroscience* 9: 221.
- Phan A, Suschkov S, Molinaro L, Reynolds K, Lymer JM, Bailey CD, Kow LM, MacLusky NJ, Pfaff DW, and Choleris E (2015) Rapid increases in immature synapses parallel estrogen-induced hippocampal learning enhancements. *Proceedings of the National Academy of Science USA* 112(52): 16018–16023.
- Rahn EJ, Guzman-Karlisson MC, and David Sweatt J (2013) Cellular, molecular, and epigenetic mechanisms in non-associative conditioning: Implications for pain and memory. *Neurobiology of Learning and Memory* 105: 133–150.
- Refolo LM, Salton SRJ, Anderson JP, Mehta P, and Robakis NK (1989) Nerve and epidermal growth factors induce the release of the Alzheimer amyloid precursor from PC 12 cells. *Biochemical and Biophysical Research Communications* 164: 664–670.
- Sweatt JD (2016) Neural plasticity and behavior - sixty years of conceptual advances. *Journal of Neurochemistry* 139 Suppl 2: 179–199.
- Woldemichael BT, Bohacek J, Gapp K, and Mansuy IM (2014) Epigenetics of memory and plasticity. *Progress in Molecular Biology and Translational Science* 122: 305–340.