



Hagfish from the Cretaceous Tethys Sea and a reconciliation of the morphological–molecular conflict in early vertebrate phylogeny

Tetsuto Miyashita^{a,b,1}, Michael I. Coates^a, Robert Farrar^c, Peter Larson^c, Phillip L. Manning^d, Roy A. Wogelius^d, Nicholas P. Edwards^{d,e}, Jennifer Anné^{d,f}, Uwe Bergmann^e, A. Richard Palmer^b, and Philip J. Currie^b

^aDepartment of Organismal Biology and Anatomy, University of Chicago, Chicago, IL 60637; ^bDepartment of Biological Sciences, University of Alberta, Edmonton T6G 2E9, Canada; ^cBlack Hills Institute of Geological Research, Hill City, SD 57745; ^dSchool of Earth and Environmental Sciences, University of Manchester, M13 9PL Manchester, United Kingdom; ^eStanford Synchrotron Radiation Lightsource, SLAC National Accelerator Laboratory, Menlo Park, CA 94025; and ^fThe Children's Museum of Indianapolis, Indianapolis, IN 46208

Edited by David M. Hillis, The University of Texas at Austin, Austin, TX, and approved December 14, 2018 (received for review August 28, 2018)

Hagfish depart so much from other fishes anatomically that they were sometimes considered not fully vertebrate. They may represent: (i) an anatomically primitive outgroup of vertebrates (the morphology-based craniate hypothesis); or (ii) an anatomically degenerate vertebrate lineage sister to lampreys (the molecular-based cyclostome hypothesis). This systematic conundrum has become a prominent case of conflict between morphology- and molecular-based phylogenies. To date, the fossil record has offered few insights to this long-branch problem or the evolutionary history of hagfish in general, because unequivocal fossil members of the group are unknown. Here, we report an unequivocal fossil hagfish from the early Late Cretaceous of Lebanon. The soft tissue anatomy includes key attributes of living hagfish: cartilages of barbels, postcranial position of branchial apparatus, and chemical traces of slime glands. This indicates that the suite of characters unique to living hagfish appeared well before Cretaceous times. This new hagfish prompted a reevaluation of morphological characters for interrelationships among jawless vertebrates. By addressing nonindependence of characters, our phylogenetic analyses recovered hagfish and lampreys in a clade of cyclostomes (congruent with the cyclostome hypothesis) using only morphological data. This new phylogeny places the fossil taxon within the hagfish crown group, and resolved other putative fossil cyclostomes to the stem of either hagfish or lamprey crown groups. These results potentially resolve the morphological–molecular conflict at the base of the Vertebrata. Thus, assessment of character nonindependence may help reconcile morphological and molecular inferences for other major discords in animal phylogeny.

Myxinoidea | cyclostome | monophyly | synchrotron | soft tissue

Hagfish and lampreys (cyclostomes) represent the only living lineages of jawless vertebrates (or agnathans) (1). Cyclostomes are generally considered primitive morphologically with respect to jawed vertebrates (gnathostomes) (1, 2). However, consensus is still lacking about cyclostome relationships. None of the traits proposed to unite hagfish and lampreys sets them apart clearly from extinct lineages of jawless vertebrates (3, 4). Furthermore, characters distribute asymmetrically. Hagfish have fewer traits than lampreys in common with gnathostomes, and are characterized by rudimentary state or absence of otherwise pan-vertebrate traits such as electroreceptors or taste buds (5, 6). Thus, resolving the cyclostome relationships holds direct implications for defining the vertebrate crown group, and for describing early character evolution in that clade. Motivated by this prospect, a number of analyses have set out to clarify positions of hagfish and lampreys with respect to gnathostomes.

The endeavor has generated variants of four main topological schemes (Fig. 1 *A–D*). Early classifications split vertebrates between agnathans and gnathostomes, with cyclostomes nested within the former (Fig. 1*A*) (7). With the advent of cladistics, this

dichotomy was challenged: hagfish fell outside the vertebrate node, rendering cyclostomes paraphyletic (craniate hypothesis; Fig. 1*B*) (8). More recent analyses of morphological data continue to support the paraphyly, but differ from the earlier analyses in placing fossil agnathans on the gnathostome stem (Fig. 1*D*) (9–16). Meanwhile, cyclostome paraphyly has been contested by molecular phylogenetics, which nearly unanimously favor cyclostomes as a clade (cyclostome hypothesis; Fig. 1*C*) (17, 18). Thus, the controversy has been locked in a stalemate between the molecular inferences for cyclostome monophyly (Fig. 1*C*) and the morphologically supported cyclostome paraphyly (Fig. 1*D*).

Despite the ever more complete molecular datasets, consensus has not been achieved. This is because (i) monophyly is an anticipated outcome of long-branch attraction in molecular data; and (ii) fossil taxa that potentially break up the long branches are only accessible via morphological data. Interpretations of character polarities are radically different between the two topologies. The morphology-supported paraphyly implies rudimentary state or absence of traits in hagfish as primitive (8). For the monophyly,

Significance

Jawless, boneless, and virtually without fossil record, hagfish have long escaped systematists' grip on their place among other fish. Yet their systematic resolution is critical to define vertebrates as a clade. Here we report an unequivocal fossil hagfish from the Cretaceous Mediterranean. Using this fossil to calibrate the evolutionary history of the group, our analysis supports hagfish and lampreys as sister groups, which likely diverged from one another in early Paleozoic times. As a result, vertebrates have a deep dichotomy, where some fossil jawless vertebrates sit closer to hagfish and lampreys than to jawed vertebrates. We showed that morphology-based analysis converged onto molecular inferences when characters are coded nonindependently, providing a case study for morphological–molecular conflicts in animal phylogeny.

Author contributions: T.M., A.R.P., and P.J.C. designed research; T.M. and M.I.C. performed research; T.M., R.F., P.L., P.L.M., R.A.W., N.P.E., J.A., and U.B. contributed new reagents/analytic tools; T.M., M.I.C., P.L.M., R.A.W., N.P.E., J.A., and U.B. analyzed data; R.F., P.L., and P.J.C. verified provenance; and T.M. wrote the paper with assistance from M.I.C., P.L.M., N.P.E., J.A., and A.R.P.

The authors declare no conflict of interest.

This article is a PNAS Direct Submission.

This open access article is distributed under [Creative Commons Attribution License 4.0 \(CC BY\)](https://creativecommons.org/licenses/by/4.0/).

Data deposition: Data relating to this article have been deposited on figshare (doi:10.6084/m9.figshare.7545002).

¹To whom correspondence should be addressed. Email: tetsuto@uchicago.edu.

This article contains supporting information online at www.pnas.org/lookup/suppl/doi:10.1073/pnas.1814794116/-DCSupplemental.

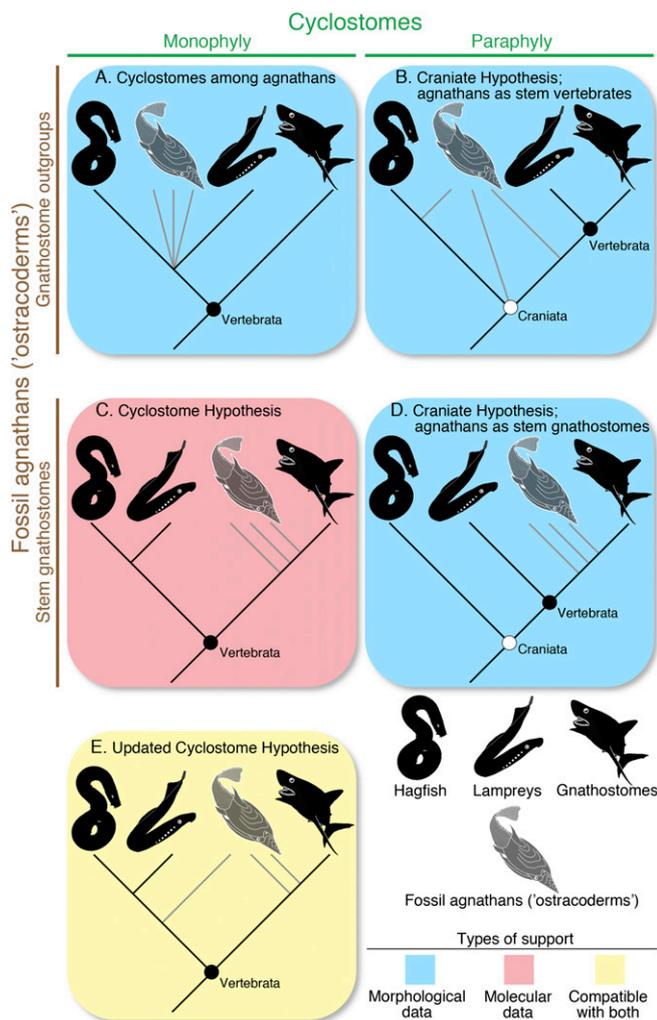


Fig. 1. A summary of hypotheses for cyclostome relationships. The hypotheses differ in cyclostome monophyly or paraphyly (columns), and about relationships of fossil “agnathans” with respect to the cyclostome and gnathostome crowns (rows). Blue box = support from morphological data; red box = support from molecular data. Yellow box = compatible with both data types. (A) Noncladistic classification schemes based on morphological data with cyclostomes nested among agnathans. (B) Craniate hypothesis in early cladistic analyses based on morphological data, where hagfish and fossil agnathans become nested outside the crown vertebrate node. (C) Cyclostome hypothesis as supported by molecular data. Fossil agnathans are assumed on the gnathostome stem. (D) Craniate hypothesis in recent analyses of morphological data, where hagfish remain nested outside the crown vertebrate node. Fossil agnathans are placed on the gnathostome stem. (E) Updated cyclostome hypothesis supported by the analyses of morphological data presented in this paper. Hagfish and lampreys form a clade of cyclostomes as supported by the molecular inferences. However, some fossil agnathans are closer to the cyclostome crown than to the gnathostome crown.

molecular trees are agnostic about character polarities, but the rudiments and absence in hagfish tend to be interpreted as secondary (2, 3). An equally parsimonious alternative—hagfish as primitive and lampreys and gnathostomes as convergent—is rarely discussed (2). As such, cyclostome relationships provide a classic example of long-branch problems hindering morphological–molecular congruence.

Although fossils are critical in reducing branch lengths, the patchy fossil record of cyclostomes remains a formidable challenge. For hagfish (myxinoidea), three soft-bodied Carboniferous forms (*Gilpichthys*, *Myxineidus*, and *Myxinikela*) have each been posited as a stem myxinoidea (19–21), but myxinoidea affinities for the former two have been questioned (9, 22, 23). *Myxinikela* is

the sole putative myxinoidea with any degree of cladistic support (9), but its similarities to living hagfish may partly be artifacts of preservation (23). As these Carboniferous forms are only preserved with a handful of morphological structures, the inferences have been based on a few myxinoidea- or cyclostome-diagnostic characters (potential keratinous teeth in *Gilpichthys*; a longitudinally elongate nasohypophyseal system in *Myxinikela*) and the general absence of lamprey-like traits (e.g., oral funnel, discrete dorsal fins) (19–21). Consequently, living hagfishes imply long ghost lineages, making it difficult to interpret the peculiar morphology of the group. Here, we report an unequivocal hagfish fossil from the Late Cretaceous of Lebanon—which reduces this gap in the fossil record by 100 My—and present an updated morphology-based phylogeny of cyclostomes. We also describe the exquisitely preserved soft-tissue anatomy of this new hagfish using trace elements mapped by synchrotron rapid-scanning X-ray fluorescence (SRS-XRF; described in *Methods* and *SI Appendix, Part B*).

Systematic Paleontology

Vertebrata Linnaeus, 1758; Cyclostomi Duméril, 1806; Myxinoidea Müller, 1834; *Tethymyxine tapirostrum* gen. et sp. nov.

Holotype. Black Hills Institute of Geological Research (BHI) 6445. A complete body fossil, exposed in lateral aspect (Fig. 2 and *SI Appendix, Figs. S1–S4*).

Etymology. The generic name is derived from “Tethys” after the Tethys Sea and “myxinos” (Latinized Greek for “slimy fish”). The specific name is derived from “tapirus” (Latinized Old Tupi for tapir) and “rostrum” (a Latin root for snout) for the elongated tapering snout.

Horizon and Locality. The type specimen was collected from sub-lithographic limestones of Hådjula Lagerstätte (Cenomanian, Upper Cretaceous), 10 km east of Byblos, Lebanon. The locality is interpreted as an interreef basin on a carbonate platform (24). See *SI Appendix, Part A* for detailed information on provenance and geological settings.

Diagnosis. *Tethymyxine tapirostrum* is a myxinoidea exhibiting the following unique combination of characters: tapering nasohypophyseal profile; tentacular cartilage not extending beyond nasohypophyseal aperture; prebranchial length nearly equal to branchial length; 8 pairs of branchial pouches; 133 slime glands on one side; and caudal fin not expanded into a round lobe.

Description and Comparisons. In visible light, BHI 6445 (Fig. 2) is preserved in full-body outline with soft tissues (body length = 313 mm). The trunk lies on its left side, whereas the head was twisted counterclockwise to expose the ventral side. The preserved tissues are marked by greater abundance of P, Mn, Fe, Cu, and Zn relative to the surrounding matrix (Fig. 2C and *SI Appendix, Fig. S3*). These tissues are inferred to be (i) composed of collagenous extracellular matrix (e.g., chondrocranium); (ii) enclosed by a highly vascularized epithelial sheet (e.g., branchial pouches, livers); or (iii) secreted keratin (e.g., infillings of slime glands, tooth-like apparatus). The original outline of the body is clearly demarcated from the matrix in most elements mapped by SRS-XRF (Fig. 2C and *SI Appendix, Fig. S3*). Al, P, S, Mn, Fe, Cu, Zn, and As are markedly more abundant in the preserved tissues than in the matrix, whereas the reverse is true for Si, Ca, and Hg. The preservation of structures and imprints is partly obscured by glue and paint, but the fossilized tissues can be distinguished from artifacts. Optically, the structural preservation is distinguished by surface topography under the artifacts. In SRS-XRF, the artifacts are clearly differentiated from the fossil and matrix based on distributions of S, Ti, V, and Fe. In particular, X-ray absorption spectroscopy reveals differential distributions of sulfur species in this specimen (*SI Appendix, Fig. S4*). Sulfate is the dominant species in all residues; organic sulfur is present in both the fossil and matrix, but virtually absent in the glue (*SI*

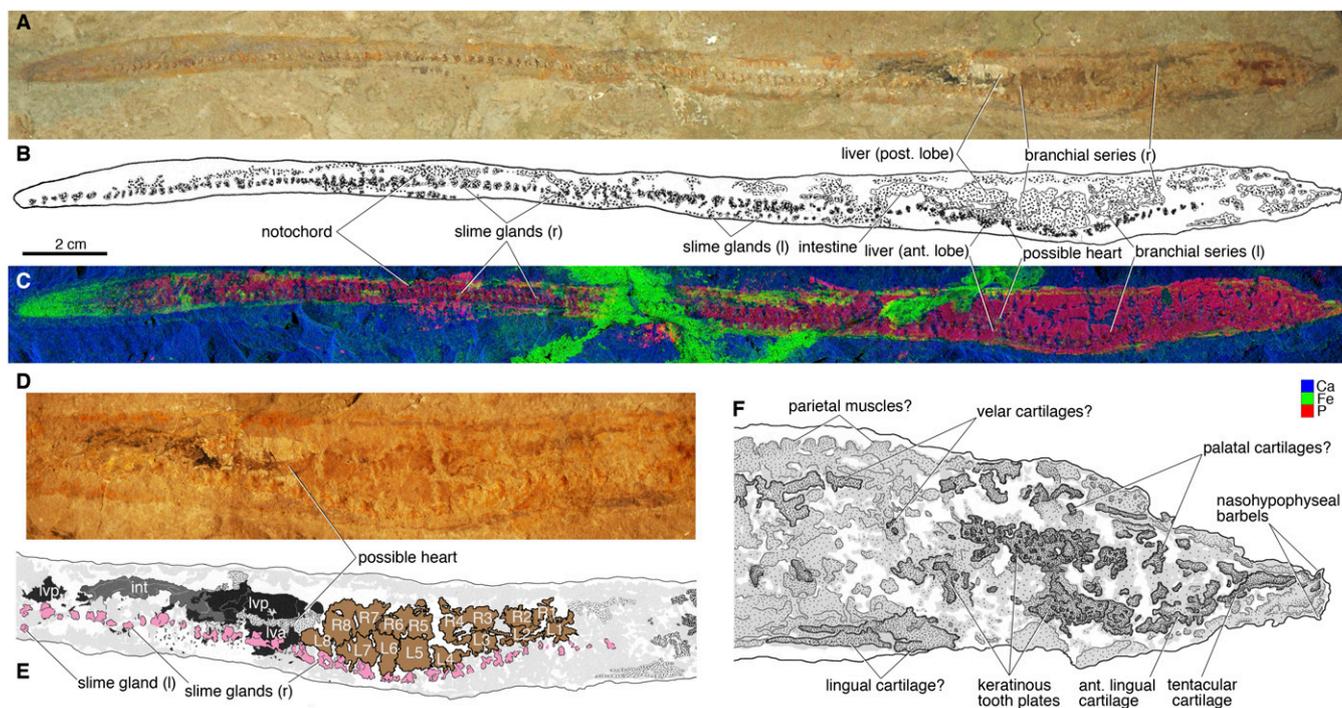


Fig. 2. *Tethymyxine tapirostrum* gen. et sp. nov, a fossil hagfish from the Cenomanian of Lebanon. Holotype (BHI 6445) in right lateral view: (A) photograph; (B) interpretive drawing; (C) false-color composite of distributions of three selected chemical elements (blue = Ca; green = Fe; red = P; Ca and Fe in HZ setup and P in LZ setup, 99.9% threshold) from SRS-XRF. The visceral anatomy of BHI 6445 in composite photograph (D) and interpretive drawing (E) in the following color codes: black = liver lobes; brown = branchial pouches; dark gray = intestine; light gray = preserved amorphous tissues; pink = slime glands; stippled gray = other soft tissues that are preserved with distinct outlines. The cranial anatomy of BHI 6445 in interpretive drawing (F) in which preserved structures are indicated in gray shades and stipples. Abbreviations: ant, anterior; int, intestine; L, left branchial pouch; l, left side; lva, liver, anterior lobe; lvp, liver, posterior lobe; post, posterior; R, right branchial pouch; r, right side.

Appendix, Part B). This result highlights the utility of SRS-XRF in identifying curatorial artifacts in fossil specimens.

The preservation of decay-prone soft tissues such as branchial pouches and slime (Fig. 2) suggests that BHI 6445 was rapidly buried (25). Although the mode of preservation is similar to that of the Early Cretaceous freshwater lamprey *Mesomyzon* from the Jehol Group of China, internal structures of BHI 6445 are not obscured by the preserved integument and muscles as in specimens of *Mesomyzon* (26) (for detailed discussion, see *SI Appendix, Part A*).

The overall characteristics clearly reject nonmyxinoïd hypotheses for the identity of BHI 6445. The absence of a mineralized skeleton, paired fins, and jaws rules out anguilliform gnathostomes like the coeval eel *Luenchelys* (27). BHI 6445 is not a lamprey either, because it does not have an oral sucker, tectal cartilages, branchial basket, dorsal fins, and other skeletal and proportional features of lampreys that would have been present given the state of preservation (25). Instead, BHI 6445 has characters diagnostic of myxinoïds, including an elongate nasohypophyseal portion of the chondrocranium, posteriorly placed branchial pouches, and slime glands.

Tethymyxine has a small head relative to its body size. The prebranchial length of BHI 6445 is shorter than that in living hagfishes, relative to the total body length (13% versus 20–30%) (28). As a consequence, the first branchial pouch sits more anteriorly than in living hagfish species, leaving a relatively short space for the lingual apparatus to occupy. The snout tapers toward the nasohypophyseal aperture (nostril). The nasohypophyseal barbels are preserved with the sigmoidal tentacular cartilage on the left side of BHI 6445. The barbels sit behind the aperture as in living *Rubicundus* (29) (Fig. 3 B and C), but the specimen differs from it in lacking a protruded nasohypophyseal tube. The tentacular cartilage is fused posteriorly with the lateral element of the anterior lingual cartilages. Both the left and right parts of

the keratinous tooth plates are preserved in association with the anterior lingual cartilages. Although several cusps can be identified by the distributions of Ca, Mn, Fe, Ni, and Hg (*SI Appendix, Fig. S3*), precise cusp number and shape cannot be determined. Unlike the preservation of comb-like tooth plates in *Gilpichthys* from Mazon Creek (20), the plates may have been pyrolysed in BHI 6445 as seen in the scales of actinopterygians from the same locality (27). Presumably the missing counterpart of BHI 6445 has the rest of preserved chondrocranial elements.

Tethymyxine has eight pairs of branchial pouches. This number is greater than in *Rubicundus* spp. ($n = 5$) and most other living hagfishes ($n = 4\sim 7$), equal to some species of *Eptatretus* and *Nemamyxine*, and smaller than in some species of *Eptatretus* ($n = 14$ in *E. polytrema*) (28). A structure immediately behind the branchial series is topographically and morphologically consistent with a heart, which has Fe in higher abundance than in the surrounding tissues (Fig. 2 D and E). The intestine extends between the anterior and posterior liver lobes. These visceral tissues have different textures and colors that allow clear delineation under natural light (*SI Appendix, Fig. S2*). However, the element mapping by SRS-XRF shows similar compositions among these tissues. P, S, and Cl appear to have greater abundances in the branchial pouches than in the intestine, and Ca seems to be the reverse (*SI Appendix, Fig. S3*).

A characteristic unique to hagfish, slime glands are preserved in an uninterrupted series of infillings that extends nearly the entire length of the body on the ventral side. The infillings are distinguished from other tissues by surface profile, high Ca, P, and S abundances, and higher levels of Cl, Ti, Mn, Fe, Cu, Ni, Zn, and Hg relative to the matrix (Fig. 2 and *SI Appendix, Figs. S2 and S3 and Table S1*). The chemical mapping is consistent with the fact that a major component of hagfish slime is tightly coiled, mucin-coated α -keratin threads (30)—pyrolysed of which are characterized by calcium phosphatic content (31). The number of the slime glands ($n = 133$

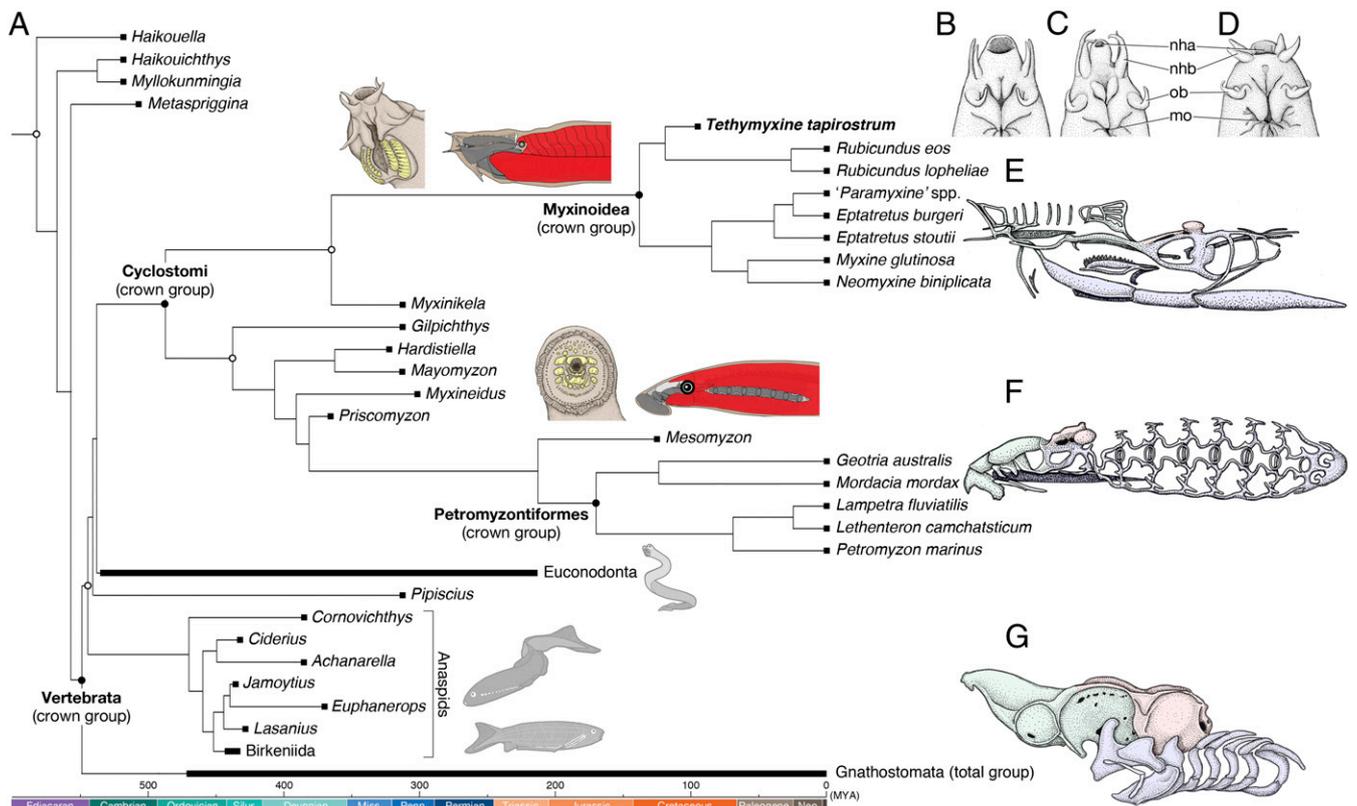


Fig. 3. A time-scaled phylogenetic tree of cyclostomes. (A) Summary tree showing cyclostome relationships. Maximum parsimony and Bayesian inferences converged onto each other in placing *Tethymyxine* within the hagfish crown group and supporting cyclostome monophyly. The precise topology is from the maximum parsimony analysis (SI Appendix, Fig. S6). Node ages represent median of 95% HPD distribution in a Bayesian molecular clock analysis of mitogenomic sequences (16S and COI) under fossilized birth–death model (see SI Appendix, Fig. S9 for a fully calibrated maximum clade credibility tree with node intervals). The crown group of cyclostomes is united by at least two morphological characters (shown on each stem): keratinous tooth plates (yellow) and periocular position of trunk muscles (red). At Top Right, the nasohypophyseal profiles are compared in ventral view among three selected crown-group hagfishes (B: *Tethymyxine tapirostrum*; C: *Rubicundus eos*; D: *Eptatretus stoutii*; the latter two based on ref. 29). To show morphological divergence among the three major crown groups of living vertebrates (E, myxinooids; F, petromyzontiforms; G, gnathostomes), each is accompanied by a chondrocranium in left lateral view (green: neural crest-derived nasohypophyseal skeleton; red: mesodermally derived neurocranium; blue: neural crest-derived pharyngeal skeleton). Filled squares represent occurrences of the terminal taxa. Crown nodes are each indicated by a filled circle, and total nodes by an empty circle. Abbreviations: nha, nasohypophyseal aperture; nhb, nasohypophyseal barbels; mo, mouth; ob, oral barbels.

on the right) is greater than in most living hagfishes ($n = 70\sim 110$), but close to that of *R. eos* ($n = 128\sim 130$) (28). The end of the tail is obscured by paint, but it is possible to outline the body by relatively high concentrations of Cu, P, S, and Fe (contained in paint, but higher in preserved tissues) and relatively low concentrations of Ca (Fig. 2C and SI Appendix, Figs. S3 and S4 and Table S1). The notochord is difficult to identify optically (as it is generally in fossils of soft-bodied vertebrates), but can be delineated with the high P and S concentrations along the dorsal midline (SI Appendix, Figs. S3 and S4 B and C). The tail tapers rather than expanding into a round lobe.

Phylogenetics. We generated a morphological dataset to test the relationships of *Tethymyxine* and cyclostomes. *Tethymyxine* has myxinooid apomorphies not identified in other fossil taxa. These traits include the posteriorly placed branchial series and slime glands, which are absent, missing, or ambiguous in *Myxinikela* and other putative fossil cyclostomes (9). To code these unequivocal and potential myxinooids in a phylogenetic dataset, we reevaluated morphological characters that are nonindependent, and enhanced contingency coding to reduce implicit weighting (32). For example, attributes of a mineralized skeleton such as bones and scales are not simply absent in the soft-bodied myxinooids. Those characters are inapplicable in this lineage. Applying no character values to attributes of biomineralization in myxinooids removes additive weight of lacking mineralization in every coded skeletal

element. This contingency coding eliminated, on average, 70% of character information from nonvertebrate outgroups and 32% from cyclostomes, which would otherwise be available to coding (SI Appendix, Parts B–D and Fig. S5 for comparative analysis).

Searches of this dataset, using both maximum parsimony and Bayesian methods, resolved *Tethymyxine* within the myxinooid crown group, sister to species of the living genus *Rubicundus* (Fig. 3A and SI Appendix, Figs. S6–S8). These two genera with an elongate snout, indented barbels, and a large number of slime glands (>100) (Fig. 3B and C) were nested outside the clade of more conventional living myxinooids (*Eptatretus*, *Myxine*, and *Neomyxine*) (Fig. 3D). *Myxinikela* was resolved as a stem myxinooid, whereas other Carboniferous forms (*Gilpichthys*, *Hardistiella*, *Mayomyzon*, and *Myxineidus*) and *Priscoomyzon* were found along the stem of lampreys. Under maximum parsimony, the crown group of cyclostomes is united by two unambiguous character changes: keratinous tooth plates and migration of postotic myomeres to position of eyes (Fig. 3A and SI Appendix, Table S3). Euconodonts, *Pipiscius*, and anaspids fell into the cyclostome stem. Together, these lineages comprised the total group of cyclostomes. Controversial Cambrian forms (*Haikouella*, *Haikouichthys*, *Metaspriggina*, and *Myllokunmingia*) were resolved in the vertebrate stem.

Given the topology obtained in the maximum parsimony analysis, and using the fossil-calibrated dataset of mitogenomic sequences, tip-dated Bayesian estimates of divergence placed the crown

of cyclostomes, the new fossil hagfish *Tethymyxine* represents an important calibration point not just for the divergence of the myxinoidean crown, but also for many morphological traits (e.g., slime glands) that set hagfish apart from all other vertebrates. The morphologically supported Cyclostomi implies some of the characters conventionally considered primitive in hagfish (e.g., the absence of extraocular muscles, electroreceptors, or taste buds) as specialized. This view is consistent with the developmental features of hagfish interpreted as vestigial and derived (e.g., axial skeleton) (38, 41). Between two living cyclostome lineages, lampreys may serve as a better outgroup to gnathostomes, but a full extent of their utility as an outgroup remains unknown until evolutionary origins of the peculiar life history (distinct larval phase with dramatic metamorphosis) are resolved (42, 43).

Methods

SRS-XRF was performed at beam line 6–2 at the Stanford Synchrotron Radiation Lightsource (SSRL), SLAC National Accelerator Laboratory. SRS-XRF is a noninvasive technique that can spatially resolve chemical element composition in large samples in short time frames (44). For the new phylogeny, three lines of analyses were performed: (i) maximum parsimony analyses; (ii) nonclock Bayesian analyses; and (iii) clock analyses. The morphological dataset was the same for all of the analyses and included 52 taxa (SI Appendix, Table S2) and 168 characters (SI Appendix, Part E). For the parsimony analyses, heuristic search was used with no topological constraint. For the

non-clock-based Bayesian analyses, each analysis was run for 5 million generations with 2 runs of 4 chains, without topological constraint, and sampling every 5,000 generations. Finally, we generated a dataset with 16S and COI sequences for extant taxa and with fossil calibrations. This dataset was subjected to a Bayesian analysis under the fossilized birth–death model, given topology obtained from the parsimony analysis. This analysis provided divergence estimates shown in SI Appendix, Fig. S9. For full description, see SI Appendix, Parts B–E. Original data for SRS-XRF and phylogenetic analyses are placed in a permanent depository (45).

ACKNOWLEDGMENTS. We thank E. Stanley for providing scans of *Rubicundus*; J.-B. Caron, G. Clément, J. Hurum, P. Janvier, Z. Johanson, O. Matton, T. Mörs, M. Purnell, T. Schossleitner, W. Simpson, and S. Walsh for collections access; M. Burns, E. Koppelhus, and K. Miyashita for logistical support; the Executive Committee (2017) of the Society of Vertebrate Paleontology for advice and recognition regarding depository of BHI 6445. This study was supported by National Science Foundation Grants 0917922 and 1541491 (to M.I.C.), National Engineering and Sciences Research Council grants (RGPIN 04863 and RGPAS 462299 to A.R.P., RGPIN 04715 to P.J.C.), and Science and Technology Facilities Council Grant ST/M001814/1 (to P.L.M.). Use of the Stanford Synchrotron Radiation Lightsource is supported by the U.S. Department of Energy (DOE), Office of Science, Office of Basic Energy Sciences under Contract DE-AC02-76SF00515. The SSRL Structural Molecular Biology Program is supported by the DOE Office of Biological and Environmental Research, and by the National Institutes of Health (NIH), National Institute of General Medical Sciences (NIGMS) (including P41GM103393). The contents of this publication are solely the responsibility of the authors and do not necessarily represent the official views of NIGMS or NIH.

- Janvier P (1996) *Early Vertebrates* (Clarendon Press, Oxford).
- Janvier P (2007) Homologies and evolutionary transitions in early vertebrate history. *Major Transitions in Vertebrate Evolution*, eds Anderson JS, Sues H-D (Indiana Univ Press, Bloomington), pp 57–121.
- Hardisty MW (1982) Lampreys and hagfishes: Analysis of cyclostome relationships. *The Biology of Lampreys*, eds Hardisty MW, Potter IC (Academic, New York), pp 165–259.
- Yalden DW (1985) Feeding mechanisms as evidence for cyclostome monophyly. *Zool J Linn Soc* 84:291–300.
- Braun CB, Northcutt RG (1997) The lateral line system of hagfishes (Craniata: Myxinoidea). *Acta Zool* 78:247–268.
- Braun CB (1998) Schreiner organs: A new craniate chemosensory modality in hagfishes. *J Comp Neurol* 392:135–163.
- Moy-Thomas JA, Miles RS (1971) *Palaeozoic Fishes* (Chapman, London), 2nd Ed.
- Løvtrup S (1977) *The Phylogeny of the Vertebrata* (Wiley, New York).
- Gabbott SE, et al. (2016) Pigmented anatomy in Carboniferous cyclostomes and the evolution of the vertebrate eye. *Proc Biol Sci* 283:20161151.
- Janvier P (1996) The dawn of vertebrates: Characters versus common ascent in the rise of current vertebrate phylogenies. *Palaeontology* 39:259–287.
- Donoghue PCJ, Forey PL, Aldridge RJ (2000) Conodont affinity and chordate phylogeny. *Biol Rev Camb Philos Soc* 75:191–251.
- Gess RW, Coates MJ, Rubidge BS (2006) A lamprey from the Devonian period of South Africa. *Nature* 443:981–984.
- Khonsari RH, Li B, Vernier P, Northcutt RG, Janvier P (2009) Agnathan brain anatomy and craniate phylogeny. *Acta Zool* 90:52–68.
- Sansom RS, Freedman K, Gabbott SE, Aldridge RJ, Purnell MA (2010) Taphonomy and affinity of an enigmatic Silurian vertebrate, *Jamoytius kerwoodi* White. *Palaeontology* 53:1393–1409.
- Morris SC, Caron J-B (2014) A primitive fish from the Cambrian of North America. *Nature* 512:419–422.
- Turner S, et al. (2010) False teeth: Conodont-vertebrate phylogenetic relationships revisited. *Geodiversitas* 32:545–594.
- Heimberg AM, Cowper-Sallari R, Sémon M, Donoghue PC, Peterson KJ (2010) MicroRNAs reveal the interrelationships of hagfish, lampreys, and gnathostomes and the nature of the ancestral vertebrate. *Proc Natl Acad Sci USA* 107:19379–19383.
- Near TJ (2009) Conflict and resolution between phylogenies inferred from molecular and phenotypic data sets for hagfish, lampreys, and gnathostomes. *J Exp Zool B Mol Dev Evol* 312:749–761.
- Bardack D (1991) First fossil hagfish (myxinoidea): A record from the pennsylvanian of Illinois. *Science* 254:701–703.
- Bardack D, Richardson ES (1977) New agnathan fishes from the Pennsylvanian of Illinois. *Fieldiana Geol* 33:489–510.
- Poplin C, Sotty D, Janvier P (2001) Un Myxinoïde (Craniata, Hyperotreti) dans le Konservat-Lagerstätte Carbonifère supérieur de Montceau-les-Mines (Allier, France). *Comptes Rendus Académie Sci - Ser IIA - Earth. Planet Sci* 332:345–350.
- Germain D, Sanchez S, Janvier P, Tafforeau P (2014) The presumed hagfish *Myxineidus gononorum* from the Upper Carboniferous of Montceau-les-Mines (Saône-et-Loire, France): New data obtained by means of propagation phase contrast X-ray synchrotron microtomography. *Ann Paleontol* 100:131–135.
- Janvier P, Sansom RS (2016) Fossil hagfishes, fossil cyclostomes, and the lost world of “ostracoderms.” *Hagfish Biology*, eds Edwards SL, Goss GG (CRC Press, Boca Raton, FL), pp 73–93.
- Hückel U (1970) Die Fischschiefer von Haqel und Hjoula in der Oberkreide des Libanon. *Neues Jahrb Geol Palaontol Abh* 135:113–149.
- Sansom RS, Gabbott SE, Purnell MA (2013) Atlas of vertebrate decay: A visual and taphonomic guide to fossil interpretation. *Palaeontology* 56:457–474.
- Chang MM, Zhang J, Miao D (2006) A lamprey from the Cretaceous Jehol biota of China. *Nature* 441:972–974.
- Forey PL, Yi L, Patterson C, Davies CE (2003) Fossil fishes from the Cenomanian (Upper Cretaceous) of Namoura, Lebanon. *J Syst Palaeontology* 1:227–330.
- Fernholm B (1998) Hagfish systematics. *The Biology of Hagfishes*, eds Jørgensen JM, Lomholt JP, Weber RE, Malte H (Chapman, London), pp 33–44.
- Fernholm B, et al. (2013) Hagfish phylogeny and taxonomy, with description of the new genus *Rubicundus* (Craniata, Myxinoidea). *J Zool Syst Evol Res* 51:296–307.
- Winegard T, et al. (2014) Coiling and maturation of a high-performance fibre in hagfish slime gland thread cells. *Nat Commun* 5:3534.
- Saitta ET, et al. (2017) Low fossilization potential of keratin protein revealed by experimental taphonomy. *Palaeontology* 60:547–556.
- Brazeau MD (2011) Problematic character coding methods in morphology and their effects. *Biol J Linn Soc Lond* 104:489–498.
- O'Reilly JE, et al. (2016) Bayesian methods outperform parsimony but at the expense of precision in the estimation of phylogeny from discrete morphological data. *Biol Lett* 12:20160081.
- Kuraku S, Kuratani S (2006) Time scale for cyclostome evolution inferred with a phylogenetic diagnosis of hagfish and lamprey cDNA sequences. *Zool Sci* 23:1053–1064.
- Goudemand N, Orchard MJ, Urdy S, Bucher H, Tafforeau P (2011) Synchrotron-aided reconstruction of the conodont feeding apparatus and implications for the mouth of the first vertebrates. *Proc Natl Acad Sci USA* 108:8720–8724.
- Krejsa RJ, Bringas P, Slavkin HC (1990) A neontological interpretation of conodont elements based on agnathan cyclostome tooth structure, function, and development. *Lethaia* 23:359–378.
- Terrill DF, Henderson CM, Anderson JS (2018) New applications of spectroscopy techniques reveal phylogenetically significant soft tissue residue in Paleozoic conodonts. *J Anal At Spectrom* 33:992–1002.
- Ota KG, Fujimoto S, Oisi Y, Kuratani S (2011) Identification of vertebra-like elements and their possible differentiation from sclerotomes in the hagfish. *Nat Commun* 2:373.
- Carlström D (1963) A crystallographic study of vertebrate otoliths. *Biol Bull* 125:441–463.
- Langille RM, Hall BK (1993) Calcification of cartilage from the lamprey *Petromyzon marinus* (L.) in vitro. *Acta Zool* 74:31–41.
- Oisi Y, Ota KG, Kuraku S, Fujimoto S, Kuratani S (2013) Craniofacial development of hagfishes and the evolution of vertebrates. *Nature* 493:175–180.
- Miyashita T (2016) Fishing for jaws in early vertebrate evolution: A new hypothesis of mandibular confinement. *Biol Rev Camb Philos Soc* 91:611–657.
- Miyashita T, Coates MJ (2016) The embryology of hagfishes and the evolution and development of vertebrates. *Hagfish Biology*, eds Edwards SL, Goss GG (CRC Press, Boca Raton, FL), pp 95–127.
- Bergmann U, Manning PL, Wogelius RA (2012) Chemical mapping of paleontological and archeological artifacts with synchrotron X-rays. *Annu Rev Anal Chem (Palo Alto Calif)* 5:361–389.
- Miyashita, et al. (2019) Data from “Hagfish from the Cretaceous Tethys Sea and a reconciliation of the morphological–molecular conflict in early vertebrate phylogeny: Data files.” Figshare. Available at <https://figshare.com/s/9a63bc42be68a85559f7>. Deposited January 3, 2019.