Association of Ongoing Cerebral Oxygen Extraction During Deep Hypothermic Circulatory Arrest With Postoperative Brain Injury

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Cardiac surgery utilizing circulatory arrest is most commonly performed under deep hypothermia (~18°C) to suppress tissue oxygen demand and provide neuroprotection during operative circulatory arrest. Studies investigating the effects of deep hypothermic circulatory arrest (DHCA) on neurodevelopmental outcomes of patients with congenital heart disease give conflicting results. Here, we address these issues by quantifying changes in cerebral oxygen saturation, blood flow, and oxygen metabolism in neonates during DHCA and investigating the association of these changes with postoperative brain injury. Neonates with critical congenital heart disease undergoing DHCA were recruited for continuous intraoperative monitoring of cerebral oxygen saturation (ScO2) and an index of cerebral blood flow (CBF) using 2 noninvasive optical techniques, diffuse optical spectroscopy (DOS) and diffuse correlation spectroscopy (DCS). Pre- and postoperative brain magnetic resonance imaging (MRI) was performed to detect white matter injury (WMI). Fifteen neonates were studied, and 11/15 underwent brain MRI. During DHCA, ScO2 decreased exponentially in time with a median decay rate of -0.04 min⁻¹. This decay rate was highly variable between subjects. Subjects who had larger decreases in ScO2 during DHCA were more likely to have postoperative WMI (P = 0.02). Cerebral oxygen extraction Greater decreases in ScO2 during DHCA are associated with post-operative brain injury.

Central Message

Cerebral oxygen saturation decreases during deep hypothermic circulatory arrest, and the extent of this decrease in oxygenation is associated with an increased risk of postoperative brain injury.

Abbreviations: CBFi, cerebral blood flow index; CBF, cerebral blood flow; CBV, cerebral blood volume; CHD, congenital heart disease/defect; CICU, cardiac intensive care unit; CHOP, The Children’s Hospital of Philadelphia; CMRO2, cerebral metabolic rate of oxygen; CW-NIRS, continuous-wave near infrared spectroscopy; DCS, diffuse correlation spectroscopy; FD-DOS, frequency-domain diffuse optical spectroscopy; FD-NIRS, frequency-domain near infrared spectroscopy; Hb, deoxyhemoglobin, measured with DOS; HbO2, oxyhemoglobin, measured with DOS; Hct, hematocrit, measured from blood gas; Hgb, hemoglobin, measured from blood gas; HLHS, hypoplastic left heart syndrome; IQR, interquartile range; OEF, cerebral oxygen extraction fraction; SpO2, peripheral capillary oxygen saturation; ScO2, cerebral tissue oxygen saturation; WMI, white matter injury

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All procedures were approved by the Institutional Review Board (11-008191, initially approved August 08, 2011).

Parents were approached for consent after birth and prior to day of surgery.

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persists during DHCA and varies widely from patient-to-patient. Patients with a higher degree of oxygen extraction during DHCA were more likely to show new WMI in postoperative MRI. These findings suggest cerebral oxygen extraction should be monitored during DHCA to identify patients at risk for hypoxic-ischemic injury, and that current commercial cerebral oximeters may underestimate cerebral oxygen extraction.

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INTRODUCTION

Congenital heart defects (CHD) are the most common birth defect, affecting approximately 40,000 newborns each year in the United States. Nearly one third of these children require cardiac surgery during the neonatal period.1 While surgical advances over the last several decades have improved survival, neurodevelopmental disabilities remain a significant morbidity among survivors. As a result, clinical and investigative focus has shifted from survival beyond the neonatal period toward neurologic sequelae. Magnetic resonance imaging studies during the neonatal period in this population exhibit a high prevalence of a specific form of hypoxic-ischemic white matter injury (WMI), which is identical in location and MRI characteristics to the punctate injury seen in premature infants termed periventricular leukomalacia.2,3 Notably, WMI in preterm survivors has been directly linked to long-term cognitive outcomes.2 Investigations of school age survivors who had various forms of CHD reveal problems with academic achievement, fine and gross motor function, visual-spatial skills, and executive function,3 all of which are similar to cognitive outcomes seen in survivors of prematurity.3,6

Recent investigations have focused on identifying perioperative risk factors for WMI in CHD and subsequent neurodevelopmental disabilities. Although the exact timing and cause of perioperative WMI remains unknown, previous studies have identified cardiac diagnosis, brain immaturity, duration of deep hypothermic circulatory arrest (DHCA), and timing of surgery as risk factors.9–12 Specifically, in a heterogeneous cohort of 153 infants with severe CHD, Becs et al showed that risk for new or worsened postoperative WMI was highest in neonates with single ventricle physiology and was associated with brain maturity and with use and duration of DHCA.10 Other studies have not found an association between DHCA duration and WMI,11,12 and other techniques such as antegrade cerebral perfusion have not been shown to be superior to DHCA for neuroprotection.14

Overall, the published results are in conflict about the significance of DHCA on the risk of postoperative WMI. Thus, in this contribution we investigate changes in cerebral hemodynamics during DHCA and explore their association with WMI. We utilize noninvasive diffuse optical and diffuse correlation spectroscopies (DOS and DCS, respectively) to quantify cerebral oxygen saturation and blood flow continuously throughout the procedure.15–17 Specifically, we investigate these changes during DHCA to ascertain whether deep hypothermia at \(~18^\circ\text{C}\) suppresses cerebral metabolism sufficiently to provide neuroprotection during circulatory arrest and how intraoperative cerebral hemodynamics relate to the risk for postoperative WMI.

MATERIALS AND METHODS

Patient Population

The subjects reported herein were taken from a larger study cohort. All term newborns (37–42 weeks gestation) with pre- or postnatally diagnosed critical CHD admitted to the cardiac intensive care unit (CICU) at The Children’s Hospital of Philadelphia (CHOP) were screened for study inclusion. Exclusion criteria included: birth weight less than 2 kg, a history of perinatal depression (ie, 5-minute APGAR<5 or cord blood pH<7.0), perinatal seizures, evidence of end-organ injury, preoperative cardiac arrest, and significant preoperative intracerebral hemorrhage (eg, grade 3 or 4 intraventricular hemorrhage). Since we were not studying neurodevelopmental outcomes, genetic syndromes were not an exclusion. Parents were approached for consent after birth and prior to day of surgery for pre- and postoperative MRI and optical monitoring during the perioperative period. A subset of enrolled patients who underwent cardiac surgery requiring DHCA is included in this manuscript.

Study Protocol

This study protocol has been described previously.11,16,18,19 All procedures were approved by the Institutional Review Board. Patient demographic data were recorded. On the morning of surgery, all patients received general anesthesia. Subsequently, they underwent a brain MRI for preoperative injury assessment. After the MRI, a noninvasive optical probe was secured to the forehead for continuous DOS/DCS measurements of cerebral oxygenation (ScO2), oxygen extraction fraction (OEF), and cerebral blood flow (CBF). In a subset of patients, a commercial oximeter (Nonin, SenSmart Model X-100) was also secured to the forehead for continuous monitoring of relative cerebral oxygen saturation (rSO2). All patients
underwent cardiac surgery utilizing cardiopulmonary bypass with DHCA. Antegrade cerebral perfusion was not used. pH-stat blood gas management was used during cooling and while hypothermic; alpha stat was used during rewarming and at normothermia per institutional protocol. Patients were cooled to a target nasopharyngeal (NP) temperature of 18°C prior to initiating DHCA.

Approximately 1 week after surgery, patients underwent a postoperative follow-up MRI scan without general anesthesia. This scan enabled us to assess the development and/or progression of brain injury.

**DOS/DCS Measurements**

DOS and DCS utilize near-infrared light to noninvasively probe static and dynamic physiological properties of cortical brain tissue. Our custom-made optical instrument combines these 2 techniques on a mobile cart that can be used in the operating room (Video 1).

DOS (also called frequency-domain near-infrared spectroscopy [FD-NIRS]) is a widely accepted method to quantify tissue oxygenation. Multidistance frequency-domain DOS (FD-DOS), employed in this study, is capable of accurate quantification of cerebral tissue oxygen saturation (ScO2), that is, in contrast to, commercial oximeters that employ continuous-wave NIRS (CW-NIRS) to monitor trends in cerebral oxygen saturation. FD-DOS uses photon diffusion theory to relate the measured amplitude attenuation and phase shift of modulated and multiply scattered light detected on the tissue surface to determine the wavelength-dependent tissue absorption (μs) and scattering (μs') properties. These properties are determined by fitting to the semi-infinite medium solution of the photon diffusion equation for a homogeneous medium. The phase and AC amplitude (AC) of the detected light quantify the optical properties of the tissue. Specifically, the slope of the phase vs source-detector separation on the tissue surface (r) and the slope of the ln(AC × r²) vs r were determined and used to compute μs and μs', respectively. Data were discarded if these linear fits had a Pearson correlation coefficient R² < 0.975, which was usually caused by probe dislodgement during patient repositioning.

The wavelength- and time-dependent absorption coefficient, μs(λ, t), in turn, depends linearly on oxy- ([HbO2]) and deoxy-hemoglobin ([Hb]) concentration. Therefore, measurements of the tissue absorption at multiple wavelengths yield these 2 concentrations in absolute units. From [HbO2] and [Hb], we derive total hemoglobin concentration (THC=[HbO2]+[Hb]) and cerebral tissue oxygen saturation (ScO2=[HbO2]/THC). Cerebral oxygen extraction fraction (OEF) is calculated from ScO2 and the arterial oxygen saturation (SaO2) measured clinically from an arterial blood gas. Cerebral blood volume (CBV, mL/100g of tissue) can be calculated from THC. The FD-DOS device employed in the present study (Imagent, ISS Inc., Champaign, IL) is amplitude modulated at 110 MHz and employs source lasers at 2 wavelengths, λ = 688 and 830 nm.

DCS uses near-infrared light to noninvasively monitor cerebral blood flow (CBF). DCS measures the temporal fluctuations (the temporal autocorrelation function) of the light intensity at the tissue surface which is primarily caused by moving red blood cells. Although this index does not have traditional physiological units of CBF, recent studies have shown that CBF can be calibrated in absolute units, and that it correlates strongly with other gold standard measures of CBF. Jain et al, for example, validated this optically measured CBF against CBF measured in the superior sagittal sinus with phase contrast MRI in a similar population of infants with critical CHD.

Importantly, the DCS-measured CBF can be combined with FD-DOS-measured ScO2 and information about clinical arterial oxygenation measured from arterial blood samples to derive an index of the cerebral metabolic rate of oxygen consumption (CMRO2,i). Then a temperature coefficient, Q10, can be calculated to assess the temperature-dependence of metabolism during the cooling period prior to DHCA. Q10 is defined as the relative change in cerebral metabolic rate of oxygen per 10°C change in temperature:

\[
Q_{10} = \left( \frac{\text{CMRO}_{2,i}}{\text{CMRO}_{2,2}} \right)^{1/10} \tag{1}
\]

The patient interface for this hybrid FD-DOS/DCS instrument consisted of a custom-made flexible black rubber probe secured to the subject’s forehead with a soft wrap. The probe houses fiber optics for both FD-DOS and DCS. For FD-DOS, we used 4 source-detector separations (1.5, 2.0, 2.5, and 3.0 cm along the tissue surface), with 2 source fibers (ie, one for each wavelength at each separation) and 1 detection fiber.

<table>
<thead>
<tr>
<th>Table 1. Patient Demographics and Operative Parameters</th>
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<tr>
<td><strong>Demographics</strong></td>
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<td>Female, n (%)</td>
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<tr>
<td>Time-to-surgery, day</td>
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<tr>
<td>Gest. age, wk</td>
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<tr>
<td>Birth weight, kg</td>
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<td>Head circumference, cm</td>
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<tr>
<th><strong>Operative parameters</strong></th>
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<tr>
<td>Duration of cooling, min</td>
</tr>
<tr>
<td>Duration of rewarming, min</td>
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<tr>
<td>Rate of cooling, °C/min</td>
</tr>
<tr>
<td>Lowest NP temperature, °C</td>
</tr>
<tr>
<td>DHCA duration, min</td>
</tr>
<tr>
<td>Total support time, min</td>
</tr>
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</table>

Normally distributed data are reported as mean standard deviation and data that are not normally distributed are reported as median (interquartile range).
For DCS, 8 single-mode detection fibers were bundled and placed on the tissue surface 2.5 cm away from the source fiber.

RESULTS

From July 2015 to April 2017, a total of \( N = 32 \) neonates with complex CHD were recruited, and 27 of 32 underwent surgery requiring DHCA. Of these 27 subjects, \( n = 15 \) had adequate optical data quality for further analysis (see Methods). Cardiac diagnoses for these 15 subjects include: hypoplastic left heart syndrome (\( n = 9 \)) and hypoplastic or interrupted aortic arch (\( n = 6 \)). Postoperative MRI scans were not acquired for 2 patients: 1 patient was deemed medically too unstable for postoperative MRI during study period and 1 subject was withdrawn from postoperative MRI by parental request. Additionally, MRI scans were not usable for 2 patients due to motion artifacts. Therefore, MRI data were available for 11 out of the remaining 15 subjects.

Patient demographics and operative parameters are summarized in Table 1. All patients were full term with an average gestational age of 38.6 ± 0.7 weeks. Patients waited an average of 3.7 ± 1.4 days from birth until their procedure. The median (IQR) duration of DHCA was 40 (12.8) minutes. Two subjects underwent a second run of bypass and one of these 2 subjects required DHCA during their second bypass run.

Figure 1 (top) shows continuous FD-DOS and DCS measurements throughout surgery for one patient. These data demonstrate typical changes that occurred during the monitoring protocol. The concentration of cerebral oxy-hemoglobin...
Cerebral tissue oxygen saturation ($ScO_2$) during deep hypothermic circulatory arrest (DHCA) for all 15 subjects. Time axis represents time on circulatory arrest. Each dotted line represents an individual subject. The color of each dotted line represents whether new postoperative white matter injury (WMI) was (red) or was not (blue) present, with a gray dotted line representing subjects for which a postoperative MRI was not performed ($N = 4$). The solid black line is the line of best fit when fitting the data to an exponential decay. Curves are numbered with subject identifier (see Table 3). (Color version of figure is available online at http://www.semthorcardiovascsurg.com).

$\left[\text{HbO}_2\right]$, and cerebral oxygen saturation ($ScO_2$), increases after the initiation of CPB, representing a decrease in cerebral oxygen extraction fraction while the patient’s temperature is decreased to 18°C. $ScO_2$ decreases after the initiation of circulatory arrest. The cerebral blood flow index ($CBFi$) falls dramatically to zero upon initiation of circulatory arrest and increases upon reperfusion. Similar trends are observed for all 15 subjects. Figure 1 (bottom) exhibits summary changes in $ScO_2$ and $CBFi$ for all 15 patients at different time points during surgery.

$ScO_2$ decreased during DHCA in all subjects, and the magnitude of this decrease was highly variable between subjects. Figure 2 shows the trend of $ScO_2$ during DHCA for all 15 subjects. The average time on DHCA in this cohort was 41 minutes. $ScO_2$ decreases during DHCA for all subjects. Fitting $ScO_2$ vs time to an exponential decay model:

$$ScO_2 = ae^{-bt} \quad (2)$$

where $t$ is the time in minutes, yields an average initial value of $60.1 \pm 13.5$ and a median [IQR] decay constant of $-0.04 [-0.06, -0.01]$. A more negative decay constant trended toward increased risk for new WMI, but this was not statistically significant ($P = 0.11$).

The presence and volume of WMI on pre- and postoperative MRI are summarized in Table 2. Optically measured changes in cerebral oxygenation predicted postoperative WMI. Specifically, larger decreases in $ScO_2$ during DHCA predicted presence of new WMI on postoperative MRI (Figure 3, $P = 0.02$ from Wilcoxon rank-sum test, area under the receiver operating characteristic curve is 0.929 with 95% CI of 0.762—1.00).

This decrease in $ScO_2$ was not detected using commercially available cerebral oximetry. For the subset ($n = 3$) of subjects wherein a commercial oximeter was also used, rSO2 was compared with FD-DOS measured $ScO_2$ (Fig. 4). Correlation between these 2 modalities was lost during DHCA, with the commercial oximeter consistently reporting an rSO2 that was greater than FD-DOS measured $ScO_2$. In this small sample, the commercial oximeter underestimated the decrease of cerebral oxygen saturation during DHCA.

To investigate changes in oxygen metabolism with cooling, a quantitative index of cerebral metabolic rate of oxygen ($CMRO_2$) was obtained during the cooling period. Figure 5 demonstrates the dependence of $CMRO_2$ on nasopharyngeal temperature (NPT). This data yielded an average $CMRO_2$ temperature coefficient $Q_{10}$ of $1.5 \pm 0.4$. $Q_{10}$, along with the duration of cooling prior to DHCA and the lowest nasopharyngeal temperature reached during the cooling period, did not predict the degree of metabolism suppression during DHCA, as defined by the change in $ScO_2$ during DHCA ($\Delta ScO_2, DHCA$) (Fig. 6).

**DISCUSSION**

This study reports the first use of diffuse optical and correlation spectroscopies for continuous monitoring of cerebral oxygen saturation, blood flow, and oxygen metabolism during DHCA in neonates. In all 15 subjects, $ScO_2$ decreased significantly during DHCA, signifying persistent oxygen extraction at 18°C, and the magnitude of this decrease in $ScO_2$ predicted the incidence of new postoperative WMI (Figure 7). To our knowledge, this direct association between continued cerebral oxygen requirement during DHCA and postoperative brain injury has not been previously shown; however, previous studies have suggested that brain metabolism persists at 18°C.29–31

Beca et al previously reported that DHCA was positively associated with severity of postoperative WMI.10 In the present study, duration of DHCA was not significantly different between subjects with or without new postoperative WMI. Other studies, including one from our group, also did not find a significant association between duration of DHCA and postoperative WMI.11,13 This difference in findings could be due to differences in the study population (eg, DHCA used for indications other than aortic arch repair), differences in criteria for

**Table 2. Presence and Volume of White Matter Injury (WMI) on Pre- and Postoperative Magnetic Resonance Imaging**

<table>
<thead>
<tr>
<th>MRI Measured Parameters</th>
<th>Presence of preoperative WMI, n(%)</th>
<th>Volume of preoperative WMI, mm$^3$</th>
<th>Presence of new postoperative WMI, n(%)</th>
<th>Volume of new postoperative WMI, mm$^3$</th>
</tr>
</thead>
<tbody>
<tr>
<td>Presence of preoperative WMI, n(%)</td>
<td>3 (20)</td>
<td>6.3 (3.9)</td>
<td>8 (73)</td>
<td>30.3 (219.3)</td>
</tr>
</tbody>
</table>

Continuous data are reported as median (interquartile range).
defining WMI on MRI, institutional differences in operative protocols (eg, duration of cooling, DHCA and rewarming), or inclusion (or lack thereof) of possible cofounders in the prediction models. For example, we previously reported that the most significant risk factor for new postoperative WMI in a homogeneous population of infants with HLHS was time-to-surgery.11 In the present study, a larger decrease in ScO2 during DHCA, signifying greater oxygen extraction, correlated with new postoperative WMI. Since the present study was limited in the number of subjects with both usable optical data (eg, due to probe dislodgement after placement, see Methods) and postoperative MRI data, robust statistical multivariate analyses to investigate risk for new postoperative WMI could not be performed. Most likely, risk for postoperative WMI is multifactorial, and our small sample size limits the power of discovery for all but the most significant risks. For example, since time-to-surgery has been shown to correlate with lower preoperative ScO2 on day of surgery and a higher risk of postoperative WMI,11,32 it is possible that a lower baseline ScO2 could augment the effect of longer DHCA duration. Clearly, the relationship between ScO2 during DHCA and postoperative injury warrants further investigation. One subject also underwent 2 runs of DHCA, and this subject also had a large amount of new WMI on postoperative MRI. However, even when removing this subject from analysis, subjects with new postoperative WMI still had a significantly greater decrease in ScO2 during DHCA (\(P = 0.04\)).

Furthermore, we observed a range of temporal decay curves/ rates for ScO2 during deep hypothermia (Fig. 2). This variability demonstrates value for individual monitoring during DHCA, for example to determine an acceptable duration of circulatory arrest. Previous studies have also reported a range of acceptable durations for DHCA. Greeley et al investigated the effect of deep hypothermia on cerebral oxygen metabolism and reported that cerebral metabolism decreased during deep hypothermia and neuroprotection was offered in the range of 39–65 minutes.30 Similarly, Ko et al reported a range of trends in CMRO2 with temperature in neonatal swine.28 Additionally in that study, nasopharyngeal temperature was shown to be an inaccurate surrogate marker for CMRO2 based on the difference in the relationship between CMRO2 and nasopharyngeal temperature during cooling vs rewarming. The findings in that study further speak to the importance of individualized

<table>
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<tr>
<th>Subject</th>
<th>Age at Surgery (Days)</th>
<th>Presence of Preoperative WMI</th>
<th>Presence of New or Worse Postoperative WMI</th>
<th>Number of CPB Runs</th>
<th>Number of DHCA Runs</th>
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<tr>
<td>1</td>
<td>4</td>
<td>No</td>
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<tr>
<td>7</td>
<td>2</td>
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CPB, cardiopulmonary bypass; DHCA, deep hypothermic circulatory arrest; WMI, white matter injury.
cerebral oxygen metabolism monitoring intraoperatively during DHCA.

The temperature coefficient, Q10, computed in this study (Q10 = 1.5 ± 0.4) is lower than values reported in literature. Greeley et al reported an average Q10 of 3.35 (2.72, 4.17) in neonates and children, although there was a large variation in Q10 values reported in that study and no attempt to separate the effect of age was made.30 It is possible that this difference in temperature coefficients is due to the different methods used for calculating Q10. Greeley et al calculated CMRO2 from CBF measured episodically by xenon-clearance methods and the arteriovenous oxygen content difference with venous blood sampled from right jugular bulb. These measurements were performed at 5 discrete time points during cardiac surgery, and Q10 was calculated from the difference between 2 time points.30 For the temperature coefficient reported herein, CMRO2 was measured continuously, and the trend between CMRO2 and NPT was fit to an exponential model to compute Q10 (see Ko et al28). The lower temperature coefficient measured during cooling reported here may also explain the continued cerebral oxygen extraction observed during DHCA; however, Q10 was not correlated with ΔScO2,DHCA (Fig. 6).

Further studies with a larger cohort are needed to explore possible reasons for this difference in temperature coefficients, including anesthetic technique and cooling duration.

Interestingly, the decrease in ScO2 captured by the commercial oximeter was not as substantial nor prolonged as the decrease captured by the more sophisticated and accurate FD-DOS device (Fig. 4). In all 3 cases when both devices were utilized, the commercial oximeter yielded a plateau in ScO2 of around 50% while ScO2 quantified by FD-DOS continued to decrease. While believed to be accurate during normal conditions for monitoring trends, commercial oximeters likely fail, even for trend monitoring, during low blood volume states. The current generation of commercial cerebral oximetry devices does not have the ability to measure the concentration of hemoglobin in the sampled tissues as is possible with FD-DOS. In the near-infrared, the absorption of water and oxygenated hemoglobin follow the same broad trend of increased absorption with longer wavelengths, while absorption from deoxyhemoglobin decreases at longer wavelengths. Without the capacity to measure the hemoglobin content in the sampled tissue, reduction in blood volume may be incorrectly interpreted as a relative increase in oxygenated hemoglobin. Thus commercial, continuous-wave oximeters may overestimate ScO2 in low volume states such as DHCA, as reported here, and their clinical use during these cases should be limited.

A limitation in the present study was the high prevalence of poor data quality due the optical probe being dislodged after placement. Improvements to the interface of the optical probe (eg, addition of an adhesive surface) are currently underway, and a study with more subjects wherein this issue is addressed...
is needed. Additionally, cerebral oxygen metabolism is only approximately quantified in this study during DHCA, due to the failure of a simple steady-state approximation during the circulatory arrest process. However, since cerebral blood flow is negligible in circulatory arrest, it is likely that oxygen extraction fraction is a good indication for oxygen utilization and thus metabolism in this state.

CONCLUSION

Cerebral oxygen saturation decreases during DHCA, and the magnitude of this decrease predicts incidence of new postoperative WMI. This knowledge of persistent cerebral oxygen extraction under deep hypothermic conditions may provide useful information for decisions about whether to employ DHCA, and if so for how long, vs a continuous cerebral perfusion strategy. Individual monitoring of cerebral oxygen saturation during DHCA is desirable too, based on the heterogeneity of trends we have observed. Commercial NIRS oximeters may fail to accurately capture this desaturation during DHCA. Ultimately, individual differences in continued oxygen extraction need to be better understood as persistent oxygen extraction during DHCA may lead to increased risk for white matter
injury for some individuals with longer times of DHCA. These pilot results and their clinical implications merit further investigation.

Acknowledgments

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SUPPLEMENTARY MATERIAL

Scanning this QR code will take you to the article title page to access supplementary material.

APPENDIX

To enable the measurement of absolute concentrations of deoxy-hemoglobin ([Hb]) and oxy-hemoglobin ([HbO2]), FD-DOS data were also collected on a solid phantom with known optical properties prior to placing the probe on the patient’s forehead. The phantom data facilitated calculation of the coupling coefficients between optical fibers and tissue.33

FD-DOS data were analyzed using the semi-infinite solution to the photon diffusion equation for a homogeneous medium. The phase and AC amplitude (AC) of the detected light are used to quantify the optical properties of the tissue. Specifically, the slope of phase versus source-detector separation (r) and the slope of ln(AC × r^−2 ) versus r were determined and used to compute tissue absorption and scattering coefficients: μₐ, λ(λ) and μₐ, s, λ(λ), respectively.30 Data were discarded if these linear fits had a Pearson R² < 0.975. In the near infrared, tissue absorption in brain tissue is primarily a result of [HbO2], [Hb], and water:

\[ \mu_a(\lambda) = \varepsilon_{\text{HbO}_2}(\lambda) \times \left[ \text{HbO}_2 \right] + \varepsilon_{\text{Hb}}(\lambda) \times \left[ \text{Hb} \right] + 0.75 \times \mu_{a,\text{H}_2\text{O}}(\lambda) \]  

(3)

Here, ε_{\text{HbO}_2}(\lambda) and ε_{\text{Hb}}(\lambda) are the known wavelength extinction coefficients of HbO₂ and Hb, respectively, μ_{a,\text{H}_2\text{O}}(\lambda) is the absorption of pure water at wavelength \( \lambda \), and we have assumed that brain tissue volume is approximately 75% water. We reconstructed average [Hb] and [HbO2] from tissue absorption measurements at 2 wavelengths (688 nm and 826 nm) via the system of 2 equations generated by Eq. 1. From [Hb] and [HbO₂], we derived ScO₂: ScO₂ = [HbO₂] / ([Hb] + [HbO₂]) × 100%. Oxygen extraction fraction (OEF) can be calculated from ScO₂ and arterial oxygenation (SaO₂) from the following equation:

\[ \text{OEF} = \frac{\text{SaO}_2 - \text{ScO}_2}{\text{ScO}_2 - \gamma \times \text{ScO}_2} \]  

(4)

where \( \gamma \) is the percentage of blood in the venous compartment and is assumed to be 75%.

For DCS, each measured intensity autocorrelation function was fit to obtain a CBF, using the semi-infinite solution to the correlation diffusion equation for a homogeneous medium. The tissue optical properties \( \mu_a \) and \( \mu_s \) at the DCS wavelength (785 nm) are required inputs to the fit for CBF.31 Although we only measured these coefficients at 688 nm and 826 nm, we were able to derive \( \mu_a(785 \text{ nm}) \) using measured [Hb] and [HbO₂] along with the known Hb and HbO₂ extinction coefficients at 785 nm using Eq. 1. To derive \( \mu_s(785 \text{ nm}) \), we assumed a power-law wavelength dependence for \( \mu_s(\lambda) \) (ie, \( \mu_s(\lambda) = A \lambda^{-b} \)), where A and b depend on the size and number of scatterers.34

The data acquired from FD-DOS and DCS—namely, ScO₂ and CBFi—were combined to give an index of cerebral metabolic rate of oxygen (CMRO₂i) using Fick’s law:

\[ \text{CMRO}_2 = \text{OEF} \times \text{CBFi} \times \text{CaO}_2 \]  

(5)

where and CaO₂ is the arterial oxygen content which can be calculated using the formula:

\[ \text{CaO}_2 = 1.39 \times \text{SaO}_2 \times [\text{Hgb}] + 0.003 \times \text{PaO}_2 \]  

(6)

Here, 1.39 is the amount of oxygen in milliliters that a fully saturated gram of hemoglobin can carry; PaO₂ is the partial pressure of oxygen in arterial blood, which is measured from an arterial blood gas and has units of mmHg; SaO₂ is the arterial oxygen saturation; and [Hgb] is the hemoglobin concentration of arterial blood in units of grams per deciliter. CaO₂ has units of milliliters of O₂ per deciliter of blood. Equation 3 yields CMRO₂ in units of milliliters of O₂ per minute per 100 g of tissue.

REFERENCES